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Top-Down Ramanomics Instrumentation Overview: from Quantitative Ramanomics with Deep Convolutional Neural Networks for Intraoperative Point-of-Care Testing Applications to Molecular Optical Laser Examiners. Part I (Bibliographic Review)

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Abstract

This review paper provides a retrospective analysis of ramanomics technologies and their methodological predecessors, ranging from modern quantitative ramanomics using deep convolutional neural networks (used for intraoperative and point-of-care diagnostics) to the Molecular Optical Laser Examiners (MOLE) of the 1970s. The first part of the review examines the current directions of this trend, while the second part presents the achievements of the earlier period. The first review part pays the special attention to applications of ramanomics for diagnostics of "supramolecular pathologies", mechanisms of apoptosis, parabiosis, oncogenesis, redox pathologies (as well as effects of active oxygen species on cells and tissues), damages of the blood-brain barrier and neurotraumas affecting the cytoarchitectonics of the brain (or, more broadly, the architecture of neuronal connectomes). A number of works are indicated that allow us to speak about Raman analysis for spectral comparative pathological organellography of the cytoplasm. Also information is given on the integrability of ramanomics with methods of mass-spectrometric mapping of biomedical samples (i.e. RaMALDI), including for MALDI-biotyping tasks for clinical microbiology applications.

Keywords: ramanomics, qRamanomics, spectralomics, single-organelle optical omics, MALDI MS imaging, RaMALDI, simultaneous Raman and MALDI imaging, label-free time-resolved single-cell monitoring, intraoperative diagnostics; point-of-care diagnostics, convolutional neural networks.

1. Введение

Что такое раманомика?

Принципиально новым омиксным направлением в молекулярной и клеточной биологии и техническим прорывом в области биомолекулярной спектроскопии является раманомика (Kuzmin et al., 2017a). По определению из только что цитированной работы, "раманомика – ... омиксная дисциплина, использующие рамановскую микроспектрометрию при анализе биомолекулярных компонентов (в целях) молекулярного профилирования биологических структур". Аналогичное определение было институционализировано на "2nd International Symposium on Physics, Engineering and Technologies for Biomedicine" ("...Ramanomics which is a new... disciplines using Micro Raman Spectrometry with Biomolecular Component Analysis for molecular profiling of biological structures"). В настоящее время данная

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дисциплина заняла прочное место в кругу омиксных дисциплин и спектральных методов химического картирования клеток, рассматриваясь как одно из наиболее сильных и многообещающих достижений инструментальной оптики и фотоники в областях рамановской спектрометрии и биомедицинского имэджинга (Siddhanta et al., 2023).

На рубеже последних лет она перешла из разряда методов качественного имэджинга (imaging) и оценивания (estimation) распределения спектрометрируемых биомолекул в плоскости микропрепарата (нередко – фиксированного) к количественному анализу 3D распределений соответствующих биомолекул, в том числе – в ходе отклика на какие-то фармацевтические или токсикологические воздействия. Такое направление называется "quantitative Ramanomics" – "qRamanomics" (LaLone et al., 2023; Dunnington et al., 2024). Для него подходит не только обычная рамановская микроспектрометрия, но и методики спектроскопии поверхностно-усиленного комбинационного рассеяния (surface-enhanced Raman spectroscopy) субклеточного уровня разрешения (Zhang et al., 2019; Shen et al., 2021). Раманомика как способ фенотипирования и профилирования клеток позволяет выявить (для последующей идентификации и прогностики) т.н. "рамановский фенотип" (Zhang et al., 2019) – частный случай "расширенного фенотипа" или морфометрии клетки по невизуальным критериям (Jablokov, Gradov, 2016). Это важно, в частности, для разработки методов рамановской спектральной цитометрии (LaLone et al., 2019a; LaLone et al., 2019b) и оперативного (point-of-care) интраоперационного рамановского контроля метаболома пациента (DePaoli et al., 2020; Huang et al., 2023).

Раманомика позволяет регистрировать и идентифицировать спектры одиночных органелл (как "single-organelle optical omics" (Pliss et al., 2021)). Собственно, в наиболее импактном изложении данного подхода (Kuzmin et al., 2017b) акцент в названии ("Molecular profiling of single organelles for quantitative analysis of cellular heterogeneity") делался на построении карт гетерогенности клеточных структур, т.е. молекулярном профилировании органелл. А в наиболее свежем обзоре на эту тему, интегрирующем разные методы вибрационной микроспектроскопии – как рамановской, так и инфракрасной – в целях безметочного (label-free) определения и времязадерженного мониторинга биохимических конституентов клетки, акцент в дефиниции термина был сделан на субклеточном уровне ("Ramanomics has been used previously to infer the use of the technique to analyse and monitor the biochemical constituent content at a subcellular level") (Byrne, 2024). Речь идёт, в частности, о биомембранах и мембранных органеллах (как плазматической мемbrane клетки и мембранах органелл типа митохондрий, так и внеклеточных везикулах (Guerreiro et al., 2024)), а также идентификации примембранных и мембранных белков и кинетической идентификации их изменений в физиологических и цитопатологических процессах (Tian et al., 2021). Последнее направление также весьма эффективно реализуется с использованием спектроскопии поверхностно-усиленного комбинационного рассеяния (surface-enhanced Raman spectroscopy) субклеточного уровня разрешения, процитированных выше (Zhang et al., 2019; Shen et al., 2021).

Это важно для выявления клеточных патологий (конечно, не в трактовке "целлюлярной патологии" Р. Вирхова XIX века, но в также классической трактовке "молекулярной" или "супрамолекулярной патологии" (Поликар А., Бесси М., 1970).

Например, в клинической хондриомике апоптоз эндотелиальных клеток сосудов головного мозга после ряда травм может быть связан с дисфункцией пула митохондрий, а его можно идентифицировать по рамановским микроспектрам – следовательно, ряд повреждений гематоэнцефалического барьера (4B = BBBB – Blood Brain Barrier Breakdown "is a key driver of traumatic brain injury (TBI)" (Schmitt et al., 2023) может быть идентифицирован на ультраструктурном уровне с использованием локальных рамановских микроспектрометрических измерений (Schmitt et al., 2023) (без привлечения электронной-микроскопии и сравнительно-патологической органеллографии цитоплазмы (Frey-Wyssling, 1965)) – аппаратурой, ПО и БД раманомики. Аналогичное верно для выявления злокачественного перерождения тканей в процессе онкогенеза: митохондриальные изменения, соответствующие злокачественным клеткам с морфологически сложно выявляемыми изменениями, могут быть выявлены аппаратурой и цифровыми "надстройками" раманомики (Gayán et al., 2022) ("надстройка" здесь может трактоваться как "überbau", без которого концепт метода не достигает цели – аппаратура с различной

разрешающей способностью для Рамановской микроспектрометрии известна, как минимум, с 1970-х гг., но без цифровой "надстройки" для глубинного анализа данных они не могли бы дать такую диагностическую определенность; уровня автоматизации XX века не хватило бы для обеспечения мультикритериальной раманомиксной диагностики).

2. Обсуждение и результаты

Раманомика: мультипараметрическая корреляция, алгоритмы, машинное обучение.

Вполне понятно, что это было бы невозможно без специализированных алгоритмических средств. В уже цитированной работе (Kuzmin et al., 2017b) писали: "Recent developments in Raman spectroscopy instrumentation and data processing algorithms have led to the emergence of Ramanomics". По существу, рамановский микроспектрометр, используемый для работы в области раманомики, представляет собой не просто оптический прибор, а "программно-аппаратный комплекс" с набором спектральных библиотек и молекулярно-биологических баз данных, экспертными системами (как минимум) или более продвинутыми средствами глубинного анализа данных (KDD) и нейросетевой компонентой / машинным обучением, обеспечивающим пополнение баз и всё более точную идентификацию биомолекулярного контента органелл. Например, в работе того же коллектива авторов, что и в публикациях (Kuzmin et al., 2017a; Kuzmin et al., 2017b), опубликованной годом позже (Kuzmin et al., 2018), вводится алгоритм "BCAbox", усовершенствующий спектр возможностей рамановской аппаратуры (микроскоп-микроспектрометра), а в работах начала 2020-х гг. внедряется комплексный подход, основанный на использовании искусственного интеллекта/машинного обучения (Lu et al., 2020; Lawrence, 2023). Например, в только цитированной работе (Lu et al., 2020) используется свёрточная нейронная сеть (ConvNet), в то время как Лоуренс (Lawrence, 2023), критикуя производительность вычислительных подходов, используемых в "классической" раманомике, считает, что простые методы потенциально могут превзойти глубокое обучение. Он справедливо отмечает, что "хотя глубокое обучение является многообещающим, оно не обеспечивает <<квантового скачка>> в производительности" и приводит свой личный опыт, в котором "простой и интерпретируемый" метод логистической регрессии достиг точности $\approx 90,4\%$ менее, чем за минуту, а использование метода случайного леса решений – random decision forest – RDF (алгоритм машинного обучения Бреймана и Катлер, использующий ансамбль решающих деревьев и применимый для задач классификации, регрессии, кластеризации) обеспечило точность 92,6 % менее чем за 10 сек. Этот опыт противопоставляется (автором данного подхода) использованию глубоких свёрточных нейронных сетей в классических и наиболее устоявшихся методах раманомики (DCNN – Deep Convolutional Neural Networks), с помощью которых точность около 90% достигается примерно после часа обучения на современном настольном компьютере (Lawrence, 2023). Идеальным решением были бы методы машинного обучения, учитывающие биофизический контекст: "A biophysics aware machine learning method would be more welcome" (там же (Lawrence, 2023)). "Чтобы действительно знать предмет, надо охватить, изучить все его стороны, все связи и "опосредования" – гласит диалектика.

Кроме того, размерность снятых с позиционной чувствительностью данных раманомики в разы превышает как данные обычной спектроскопии, так и двумерные микрофотографии и химические карты. В цитированной статье (Gayán et al., 2022) указывается на возможность использования конфокальных микроскопов, по определению, способных не только фокусироваться в определенных точках ультраструктуры клетки, но и отстраивать трехмерные реконструкции органелл с использованием лазеров с разными длинами волн, из чего (интуитивным образом) выводится целесообразность интеграции конфокальной и рамановской идентификации в клеточной патологии. Так, в цитируемой работе (Gayán et al., 2022) пишут, что так как "раковые митохондрии демонстрируют различные профили по сравнению с нормальными в морфологии, геномном, транскриптомном, протеомном и метаболическом профиле" ("cancerous mitochondria exhibit different profiles compared with normal ones in morphology, genomic, transcriptomic, proteomic and metabolic landscape"), их дифференциальный мультипараметрический анализ на уровне одиночных клеток весьма затруднен и требует возможности анализа биомаркеров с

использованием искусственного интеллекта для комплексного анализа всего вышеперечисленного пула профилей данных ("exploring such characteristics as potential biomarkers through single-cell omics and Artificial Intelligence (AI)"). Поэтому особые надежды возлагаются на платформы для раманомики, базирующиеся на конфокальных рамановских микроспектрометрах, для идентификации соответствующих "сигнатур" в объёме клеток - на клеточном либо органеллографическом уровне ("Another study used the Ramanomics platform, which coupled confocal Raman micro-spectrometry to a biomolecular component analysis algorithm to identify signatures") ([Gayan et al., 2022](#)).

Кросс-валидируемость данных раманомики.

Существенным преимуществом раманомики как позиционно-чувствительного омиксного подхода, работающего на уровне клеток и субклеточных структур, является сравнимость/совместимость его с масс-спектрометрией, которая является движущей силой прогресса в омиксных областях, в особенности – в позиционно-чувствительных "spatiotemporal omics" ([Girolamo et al., 2013; Franceschi et al., 2013; Wolyniak et al., 2018; Sanders, Edwards, 2020; Zaikin, Borisov, 2021; Wang et al., 2022; Challen, Cramer, 2022; Pade et al., 2021](#)). Данное утверждение верно и для спектрометрии ионной подвижности ([Arthur et al., 2017; Causon et al., 2020; Bilbao et al., 2021; Delafield et al., 2022; Paglia et al., 2022](#)) (см. также некоторые диссертации на тему омиксных приложений метода спектрометрии ионной подвижности ([Donohoe, 2016; Lareau, 2016](#))). Аналогично тому, как это постулируется в раманомике, для пользователя масс-спектрометрических омик наиболее информативны не просто спектры, а пространственные распределения соединений, отраженные в форме 2D картирования с достаточным пространственным разрешением – "mass spectrometric imaging-based multi-omics" ([Belizario et al., 2015; Quanico et al., 2017; Dewez et al., 2019; Chao, Zongwei, 2021; Smets et al., 2021; Zhao et al., 2022; Wang et al. 2023a; Zhao, Cai, 2023; Wang et al., 2023b; Phulara & Seneviratne, 2024](#)). Также, как и в случае раманомики, предельным (не только в случае имэджинга, но и для всех масс-спектрометрических омик) разрешением считается анализ одиночных клеток – т.н. "single cell omics" или "single cell multi-omics" ([DeLaney et al., 2018; Lu et al., 2023; Zhao et al., 2023; Zhang et al., 2023; Zhang, Qiao, 2024](#)). При этом, в случае лазерных методов десорбции-ионизации, считается приемлемым сопряжение их с лазерной микродиссекцией клеток, как это имеет место и в рамановской спектроскопии и сопряженных с нею "омиках" ([Quanico et al., 2017; Dewez et al., 2019](#)). По сути, это есть, в методическом смысле, одно из ответвлений метода микропучковой пунктуры клетки, про которую мы недавно писали в обзоре ([Orekhov, Gradov, 2023](#)). И даже наиболее активно исследуемые объекты у раманомики (и, шире, спектраломики, включающей в себя иные вибрационно-спектрометрические омики) и масс-спектрометрических омик в достаточно существенной степени перекрываются (митохондрии ([Wang et al., 2023b](#)); нейротравмы и нейропатологии ([Mallah et al., 2023](#)); диагностика рака ([Zhang et al., 2007; Nie et al., 2016; Pralea et al., 2020; Banerjee et al., 2023](#)), включая область гепатоонкологии ([Nie et al., 2016; Lawrence, 2023](#)); и т.д.).

Поэтому, как минимум, в свете давних попыток инструментальной интеграции методов рамановской спектрометрии и MALDI-имэджинга ([Bocklitz et al., 2013; Bocklitz et al., 2015; Ryabchukov et al., 2018](#)), закончившихся возникновением подхода "RaMALDI" (который, по определению, представляет собой "simultaneous Raman and MALDI imaging"), следует полагать, что раманомика и масс-спектрометрические омики могут быть полностью интегрированы в один комплекс спектральных протоколов и комбинируемых инструментов, один из которых выдаёт на первой стадии данные неразрушающего аналитического контроля и идентификации субстанции, а второй, характеризующийся ионизацией, аблацией и десорбцией субстанции ("разрушающие"), на второй стадии эксперимента выдаёт полную качественную и количественную информацию о её составе. Этот подход применим как для живых, так и для биогенных и биокосных систем ([Skottvoll, 2022; Luo et al., 2022](#)).

Раманомика как комплементарный метод для микробиологического биотайпинга и редокс-патологии

Одним из приоритетных направлений обеспечения сопоставимости рамановского и МС картирования (в том числе – МС-имэджингового, например *MALDI MS imaging*)

является микробиология. Рамановская микроскопия или рамановская микроспектрометрия часто используются в бактериологии, микробиологии (Huang, Spiers, 2006; Mosier-Boss, 2017; Lorenz et al., 2020; Hong et al., 2021; Jian, 2023; Burioni et al., 2024), в том числе в формате рамановской цитометрии, о которой было вскользь сообщено в предшествующем разделе (Jian, 2023). Результаты рамановских измерений в микробиологии хорошо соотносятся с результатами масс-иммажинга и омиксного картирования, а также – с предварительными данными идентификации микроорганизмов методами MALDI-биотайпинга (по MALDI биотайпингу см., например: Berrazeg et al., 2013; Somboro et al., 2014; Gekenidis et al., 2014; Pranada et al., 2016; Boyer et al., 2017; Houdelet, 2015; Antonios et al., 2022; Pena et al., 2022; по корреляционному иммажингу для MALDI-биотайпинга и миеробиологического мониторинга см. наши работы: Jablokow, Gradow, 2015a; Jablokow, Gradow, 2015b; Orekhov et al., 2016; Orekhov et al., 2023; Jablokow et al., 2017; Jablokow et al. 2018; Orekhov, Gradow, 2022; Orekhov, Gradow, 2023a; Orekhov, Gradow, 2023b).

Ещё одним аспектом обеспечения сопоставимости рамановского и МС картирования, на наш взгляд, может стать окислительная модификация белков и анализ продуктов редокс-реакций в цитоплазме (в том числе, интерпретируемых в контексте насоновской теории/концепции "местной реакции протоплазмы" (Portugalov et al., 1964; Hadacek, Bachmann, 2015; Jaeken, 2017; Kosmachevskaya, Topunov, 2021; Bagatolli et al., 2021), впрочем, вполне безотносительно к его некорректным амембранистским взглядам). Известно, что МС-, равно как и МС-иммажинг являются хорошими методами *in situ* анализа окислительной модификации белков и липидов (Person et al., 2003; Cornellison et al., 2011; Bykova et al., 2011; Murray, Van Eyk, 2012; Paulech et al., 2013; Bykova, Rampitsch, 2013; Butterfield et al., 2014; Bonham et al., 2014; Lennicke et al., 2016). Можно отметить, что методы MALDI MS + FRAP and FLIP (Jablokow, Gradow, 2015a, 2015b; Orekhov et al., 2016; Orekhov et al., 2023; Jablokow et al., 2017, Jablokow et al., 2018) также являются применимыми в целях цитофизиологического и цитопатологического редокс-анализа, так как, например, метод FRAP с генетически-кодируемыми редокс-сенсорными белками, такими как HuPer, давно и широко используется для картирования внутриклеточного распределения пероксида водорода и антиоксидантных градиентов в клетках и тканях (Belousov et al., 2006; Chudakov et al., 2010; Rhee et al., 2010; Samoylenko et al., 2013; Fernandez-Garcia, Olmos, 2014; Yang, 2014; Weller et al., 2014; Jones, Sies, 2015; Quintá et al., 2016; Delfosse et al., 2016; Bilan, Belousov, 2016; Rezende et al., 2018; Brilkina et al., 2018; Asada et al., 2018; Lyublinskaya, Antunes, 2019; Smolyarova et al., 2022). Из оптических методов известен ряд методов редокс-метрической микроскопии, фиксирующих повреждения внутри клетки или же их корреляты в атмосфере и окружающей среде, возникающие под действием тех же агентов (например, методы озонометрической микроскопии (Градов, 2012; Gradow, 2013)). Раманомика же даёт возможность напрямую картировать происхождение активных форм кислорода, причём – неинвазивным и позиционно-чувствительным методом – с учётом компартментализации (Janků et al., 2019a, 2019b). В (Janků et al., 2019a) оптимистично резюмируется, что "“ramanomics” approach might provide an efficient tool of non-invasive quantitative profiling of cellular compartments and monitoring of molecular interactions" ("подход "раманомики" может обеспечить эффективный комплекс инструментов для неразрушающего-неинвазивного количественного профилирования клеточных компартментов и мониторирования молекулярных взаимодействий в них"). В работе (Janků et al., 2019b) указывается, что, кроме активных форм кислорода, можно анализировать на основе того же подхода компартментализацию окислов азота (NOx).

Раманомика как инструмент этиологического исследования в молекулярной онкологии

Примеры из вышецитированных работ (Janků et al., 2019a, 2019b) относятся к растительным клеткам, но, в действительности, ими данное направление ROS-метрии не ограничивается.

Так, например, возможно исследовать методами раманомики этиологию онкогенеза. Известно, что онкогенез нередко, в частности, связывают с:

– Накоплением активных форм кислорода (это давно известный подход, работы по которому встречаются как в старой литературе до 1990-го года (Fischer, 1987; Fischer et al.,

1987), так и в работах 1990-х гг. (Standeven, Wetterhahn, 1991; Klein, Costa, 1991; Valavanidis, 1994; Emerit, 1994; Huang et al., 1994; Shi et al., 1998; Oliński, Jurgowiak, 1999), 2000-х гг. (Nishigori et al., 2004; Schulte-Hermann et al., 2006; Okada, 2007; Panayiotidis, 2008; Marquez-Quinones, 2007; Wang, 2009), 2010-х гг. (Ralph et al., 2010; Ziech et al., 2011, 2012; Grigorov, 2012; Tamura et al., 2013; Wu, Ni, 2015; Kruk, Aboul-Enein, 2017; Moldogazieva et al., 2018; Dupuy, 2018; Medeiros, 2018; Kovacic, Abadjian, 2018; Valavanidis, 2019; Jopkiewicz, 2019), 2020-х гг. (Gokulan et al., 2020; Vostrikova et al., 2020; Okazaki, 2022; Shimura, Ushiyama, 2024));

– Электрофилами (Chouchane, 1996; Miller, 1998; MacLeod et al., 2009; Smith et al., 2014; Olsen et al., 2018; Harach et al., 2019; Gobert et al., 2021; Lei et al., 2021; Danes et al., 2021) и шире понимавшимися на ранних этапах исследований в молекулярной и клеточной онкологии зарядовыми механизмами (Cavalieri, Calvin, 1972; Andrews et al., 1979; Shkarina et al., 1984; Kovacic et al., 1986);

– Зарядом мембранны и биоэнергетикой митохондрий (Tokuoka, Morioka, 1957; Beech, 1989a; Beech, 1989b, Beech, 1994; Marino et al., 1994; Ye et al., 2011; Friday et al., 2011; Yang, Brackenbury, 2013; Gogichadze et al., 2014; Lemeshko, 2015; Forrest, 2015; Lee et al., 2016; Li et al., 2020; Nnodim, Hauwa, 2020; Gąbka et al., 2021; Kuwahara et al., 2021; Sadri et al., 2022; Skates, 2022; Begum, Shen, 2023; Delisi et al., 2024).

В то же время, рамановская (микро)спектрометрия и раманомика, используемые в анализе онкогенеза по метаболическим нарушениям (которые, в частности, могут быть связаны с воздействием редокс-факторов) (Larion et al., 2018; Lawrence, 2023), могут быть использованы для коррелирования результатов редокс-повреждений или продуктов действия активных форм кислорода и онкогенеза.

3. Заключение

Из всего вышеизложенного вполне очевидно, что внедрение методов раманомики как в молекулярную медицину, так и в молекулярную биотехнологию является приоритетной задачей, следующей из всего развития последних десятилетий.

В следующей части данной работы, планируемой к выходу в 2025 году, мы рассмотрим практический аспект инструментального развития раманомики, начиная с молекулярно-оптических лазерных анализаторов (MOLE), которые мы пытались использовать для подобных задач много лет назад.

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References

- Градов, 2012 – Градов О.В. Экспериментальные установки для озонометрической микроскопии // Медицинская техника. 2012. (6): 42-47.
- Поликар, Бесси, 1970 – Поликар А., Бесси М. Элементы патологии клетки. М.: Мир. 1970.
- Andrews et al., 1979 – Andrews E.J., Todd P.W., Kukulinsky N.E. Surface charge in foreign body carcinogenesis // Journal of Biomedical Materials Research. 1979. 13(2): 173-187. DOI: 10.1002/jbm.820130203
- Antonios et al., 2022 – Antonios M.A., Raouf M.M., Ghoniem S.A., Hassan E.H. Clinical Impact of Biotyping of Klebsiella pneumoniae Isolates From Health Care-Associated Infections Using MALDI-TOF-MS // Infectious Diseases in Clinical Practice. 2022. 30(4): e1143. DOI: 10.1097/IPC.oooooooooooo00001143
- Arthur et al., 2017 – Arthur K.L., Turner M.A., Reynolds J.C., Creaser C.S. (2017). Increasing peak capacity in nontargeted omics applications by combining full scan field asymmetric waveform ion mobility spectrometry with liquid chromatography-mass spectrometry. *Analytical chemistry*. 89(6): 3452-3459. DOI: 10.1021/acs.analchem.6b04315
- Asada et al., 2018 – Asada M., Hakimi H., Kawazu S.I. The application of the HyPer fluorescent sensor in the real-time detection of H₂O₂ in Babesia bovis merozoites in vitro // *Veterinary parasitology*. 2018. 255: 78-82. DOI: <https://doi.org/10.1016/j.vetpar.2018.03.016>
- Bagatolli et al., 2021 – Bagatolli L.A., Mangiarotti A., Stock R.P. Cellular metabolism and colloids: Realistically linking physiology and biological physical chemistry // *Progress in Biophysics and Molecular Biology*. 2021. 162: 79-88. DOI: <https://doi.org/10.1016/j.pbiomolbio.2020.06.002>

- Banerjee et al., 2023** – Banerjee S., Hatimuria M., Sarkar K., Das J., Pabbathi A., Sil P.C. Recent Contributions of Mass Spectrometry-Based “Omics” in the Studies of Breast Cancer // *Chemical Research in Toxicology*. 2023. 37(2): 137-180. DOI: <https://doi.org/10.1021/acs.chemrestox.3c00223>
- Beech, 1989a** – Beech J.A. The membrane potential theory of carcinogenesis. *Medical Hypotheses*. 1989. 29(2): 101-104. DOI: [https://doi.org/10.1016/0306-9877\(89\)90069-8](https://doi.org/10.1016/0306-9877(89)90069-8)
- Beech, 1989b** – Beech J.A. Two stage carcinogenesis by membrane potential changes // *Medical Hypotheses*. 1989. 29(3): 217-221. DOI: [https://doi.org/10.1016/0306-9877\(89\)90196-5](https://doi.org/10.1016/0306-9877(89)90196-5)
- Beech, 1994** – Beech J.A. Carcinogenesis and initiation of cell cycling by charge-induced membrane clusters may be due to mitogen receptors and Na⁺ H⁺ antiports // *Medical hypotheses*. 1994. 42(6): 385-389. DOI: [https://doi.org/10.1016/0306-9877\(94\)90158-9](https://doi.org/10.1016/0306-9877(94)90158-9)
- Begum, Shen, 2023** – Begum H.M., Shen K. Intracellular and microenvironmental regulation of mitochondrial membrane potential in cancer cells // *WIREs mechanisms of disease*. 2023. 15(3): e1595. DOI: <https://doi.org/10.1002/wsbm.1595>
- Belizario et al., 2015** – Belizario J.E., Sangiuliano B.A., Perez-Sosa M., Santos B.V., Machado-Santelli G.M. (2015). Advances in the integration of optical and mass spectrometry molecular imaging technologies: from omics data to molecular signature discovery // *Discovery Medicine*. 2015. 20(112): 393-401.
- Belousov et al., 2006** – Belousov V.V., Fradkov A.F., Lukyanov K.A., Staroverov D.B., Shakhbazov K.S., Terskikh A.V., Lukyanov S. Genetically encoded fluorescent indicator for intracellular hydrogen peroxide // *Nature methods*. 2006. 3(4): 281-286. DOI: <https://doi.org/10.1038/nmeth866>
- Berrazeg et al., 2013** – Berrazeg M., Diene S. M., Drissi M., Kempf M., Richet H., Landraud L., Rolain J.M. Biotyping of multidrug-resistant *Klebsiella pneumoniae* clinical isolates from France and Algeria using MALDI-TOF MS // *PLoS one*. 2013. 8(4): e61428. DOI: <https://doi.org/10.1371/journal.pone.0061428>
- Bilan, Belousov, 2016** – Bilan D.S., Belousov V.V. HyPer family probes: state of the art // *Antioxidants & redox signaling*. 2016. 24(13): 731-751. DOI: <https://doi.org/10.1089/ars.2015.6586>
- Bilbao et al., 2021** – Bilbao A., Gibbons B.C., Stow S.M., Kyle J.E., Bloodsworth K.J., Payne S.H., Smith R.D., Ibrahim Y.M., Baker E.S., Fjeldsted J.C. A preprocessing tool for enhanced ion mobility–mass spectrometry-based omics workflows // *Journal of proteome research*. 2021. 21(3): 798-807. DOI: <https://doi.org/10.1021/acs.jproteome.1c00425>
- Bocklitz et al., 2013** – Bocklitz T.W., Crecelius A.C., Matthaus C., Tarcea N., von Eggeling F., Schmitt M., Schubert U.S., Popp J. Deeper understanding of biological tissue: quantitative correlation of MALDI-TOF and Raman imaging // *Analytical chemistry*. 2013. 85(22): 10829-10834. DOI: <https://doi.org/10.1021/ac402175c>
- Bocklitz et al., 2015** – Bocklitz T., Bräutigam K., Urbanek A., Hoffmann F., von Eggeling F., Ernst G., Schmitt M., Schubert U., Guntinas-Lichius O., Popp J. (2015). Novel workflow for combining Raman spectroscopy and MALDI-MSI for tissue based studies // *Analytical and bioanalytical chemistry*. 2015. 407: 7865-7873. DOI: <https://doi.org/10.1007/s00216-015-8987-5>
- Bonham et al., 2014** – Bonham C.A., Steevensz A.J., Geng Q., Vacratsis P.O. Investigating redox regulation of protein tyrosine phosphatases using low pH thiol labeling and enrichment strategies coupled to MALDI-TOF mass spectrometry // *Methods*. 2014. 65(2): 190-200. DOI: <https://doi.org/10.1016/j.ymeth.2013.08.014>
- Boyer et al., 2017** – Boyer P.H., Boulanger N., Nebbak A., Collin E., Jaulhac B., Almeras L. (2017). Assessment of MALDI-TOF MS biotyping for *Borrelia burgdorferi* sl detection in *Ixodes ricinus*. *PLoS One*. 2017. 12(9): e0185430. DOI: <https://doi.org/10.1371/journal.pone.0185430>
- Brilkina et al., 2018** – Brilkina A.A., Peskova N.N., Dudenkova V.V., Gorokhova A.A., Sokolova E.A., Balalaeva I.V. Monitoring of hydrogen peroxide production under photodynamic treatment using protein sensor HyPer // *Journal of Photochemistry and Photobiology B: Biology*. 2018. 178: 296-301. DOI: <https://doi.org/10.1016/j.jphotobiol.2017.11.020>
- Burioni et al., 2024** – Burioni R., Silvestrini L., D’Orto B., Tetè G., Nagni M., Polizzi E., Gherlone E.F. Could Dental Material Reuse Play a Significant Role in Preservation of Raw Materials, Water, Energy, and Costs? Microbiological Analysis of New versus Reused Healing Abutments: An In Vitro Study. *Bioengineering*. 2024. 11(4): 387. DOI: <https://doi.org/10.3390/bioengineering11040387>

- Butterfield et al., 2014** – Butterfield D.A., Gu L., Domenico F.D., Robinson R.A. (2014). Mass spectrometry and redox proteomics: applications in disease. *Mass spectrometry reviews*. 2014. 33(4): 277-301. DOI: <https://doi.org/10.1002/mas.21374>
- Bykova, Rampitsch, 2013** – Bykova N.V., Rampitsch C. Modulating protein function through reversible oxidation: redox-mediated processes in plants revealed through proteomics. *Proteomics*. 2013. 13(3-4): 579-596. DOI: <https://doi.org/10.1002/pmic.201200270>
- Bykova et al., 2011** – Bykova N.V., Hoehn B., Rampitsch C., Banks T., Stebbing J.A., Fan T., Knox R. Redox-sensitive proteome and antioxidant strategies in wheat seed dormancy control // *Proteomics*. 2011. 11(5): 865-882. DOI: <https://doi.org/10.1002/pmic.200900810>
- Byrne, 2024** – Byrne H.J. Spectralomics—Towards a holistic adaptation of label free spectroscopy // *Vibrational Spectroscopy*. 2024. 132: 103671. DOI: <https://doi.org/10.1016/j.vibspec.2024.103671>
- Causon et al., 2020** – Causon T.J., Kurulugama R.T., Hann S. Drift-tube ion mobility-mass spectrometry for nontargeted' omics // *Methods in Molecular Biology*. 2020. 2084: 79-94. DOI: https://doi.org/10.1007/978-1-0716-0030-6_4
- Cavalieri, Calvin, 1972** – Cavalieri E., Calvin M. Charge localization in carbonium-ion of methylbenzanthracenes-clue to their mechanism of carcinogenesis // *Proceedings of the American Association for Cancer Research*. 1972. 13(MAR): 125.
- Challen, Cramer, 2022** – Challen B., Cramer R. Advances in ionisation techniques for mass spectrometry-based omics research // *Proteomics*. 2022. 22(15-16): 2100394. DOI: <https://doi.org/10.1002/pmic.202100394>
- Chao, Zongwei, 2021** – Chao Z., Zongwei C. (2021). Mass spectrometry imaging and omics for environmental toxicology research // *Progress in chemistry*. 2021. 33(4): 503. DOI: <https://doi.org/10.1016/j.cbpa.2022.102199>
- Chouchane, 1996** – Chouchane S. (1996). Etude des proprietes electrophiles generees en milieu aqueux aere, par des poussières d'hématite ou de magnétite. Relation probable avec le cancer bronchopulmonaire (Doctoral dissertation, Paris 6). [Electronic resource]. URL: <https://theses.fr/1992PA066268>
- Chudakov et al., 2010** – Chudakov D.M., Matz M.V., Lukyanov S., Lukyanov K.A. Fluorescent proteins and their applications in imaging living cells and tissues // *Physiological reviews*. 2010. 90(3): 1103-1163. DOI: <https://doi.org/10.1152/physrev.00038.2009>
- Cornellison et al., 2011** – Cornellison C.D., Dyer J.M., Plowman J.E., Krsinic G.L., Clerens S. (2011). MALDI-MS redox lipidomics applied to human hair: A first look. *International Journal of Trichology*. 2011. 3(1): 25-27. DOI: <https://doi.org/10.4103/0974-7753.82127>
- Danes et al., 2021** – Danes J.M., Palma F.R., Bonini M.G. Arsenic and other metals as phenotype driving electrophiles in carcinogenesis // *Seminars in Cancer Biology*. 2021. 76: 287-291. DOI: <https://doi.org/10.1016/j.semcan.2021.09.012>
- Delafield et al., 2022** – Delafield D.G., Lu G., Kaminsky C.J., Li L. High-end ion mobility mass spectrometry: A current review of analytical capacity in omics applications and structural investigations // *TrAC Trends in Analytical Chemistry*. 2022. 157: 116761. DOI: <https://doi.org/10.1016/j.trac.2022.116761>
- DeLaney et al., 2018** – DeLaney K., Sauer C.S., Vu N.Q., Li L. Recent advances and new perspectives in capillary electrophoresis-mass spectrometry for single cell “omics” // *Molecules*. 2018. 24(1): 42. DOI: <https://doi.org/10.3390/molecules24010042>
- Delfosse et al., 2016** – Delfosse K., Wozny M.R., Jaipargas E.A., Barton K.A., Anderson C., Mathur J. Fluorescent protein aided insights on plastids and their extensions: a critical appraisal // *Frontiers in Plant Science*. 2016. 6: 1253. DOI: <https://doi.org/10.3389/fpls.2015.01253>
- Delisi et al., 2024** – Delisi D., Eskandari N., Gentile S. Membrane potential: A new hallmark of cancer // *Advances in cancer research*. 2024. 164: 93-110. DOI: <https://doi.org/10.1016/bs.acr.2024.04.010>
- DePaoli et al., 2020** – DePaoli D., Lemoine É., Ember K., Parent M., Prud'homme M., Cantin L., Petrecca K., Leblond F., Côté D.C. Rise of Raman spectroscopy in neurosurgery: a review // *Journal of biomedical optics*. 2020. 25(5): 050901. DOI: <https://doi.org/10.1117/1.jbo.25.5.050901>
- Dewez et al., 2019** – Dewez F., Martin-Lorenzo M., Herfs M., Baiwir D., Mazzucchelli G., De Pauw E., Heeren R.M., Balluff B. Precise co-registration of mass spectrometry imaging, histology, and laser microdissection-based omics // *Analytical and bioanalytical chemistry*. 2019. 411: 5647-5653. DOI: <https://doi.org/10.1007/s00216-019-01983-z>

Donohoe, 2016 – *Donohoe G.C.* Expanding the Applications of Ion Mobility Spectrometry and Mass Spectrometry in Integrative'Omics Analyses. (PhD Dissertation, West Virginia University). 2016. DOI: <https://doi.org/10.33915/etd.5505>

Dunnington et al., 2024 – *Dunnington E.L., Wong B.S., Fu D.* Innovative Approaches for Drug Discovery: Quantifying Drug Distribution and Response with Raman Imaging // *Analytical Chemistry*. 2024. 96(20): 7926-7944. DOI: <https://doi.org/10.1021/acs.analchem.4co1413>

Emerit, 1994 – *Emerit I.* Reactive oxygen species, chromosome mutation, and cancer: possible role of clastogenic factors in carcinogenesis // *Free Radical Biology and Medicine*. 1994. 16(1): 99-109. DOI: [https://doi.org/10.1016/0891-5849\(94\)90246-1](https://doi.org/10.1016/0891-5849(94)90246-1)

Fernandez-Garcia, Olmos, 2014 – *Fernandez-Garcia N., Olmos E.* ROS and NOS imaging using microscopical techniques // *Plant Image Analysis: Fundamentals and Applications*. 2014. 245. DOI: <http://dx.doi.org/10.1201/b17441-13>

Fischer et al., 1987 – *Fischer S.M., Cameron G.S., Baldwin J.K., Jascheway D.W., Patrick, K.E.* Reactive oxygen in the tumor promotion stage of skin carcinogenesis // *Lipids*. 1987. 23(6): 592-597. DOI: <https://doi.org/10.1007/bf02535603>

Fischer, 1987 – *Fischer S.* Reactive oxygen in the tumor promotion stage of carcinogenesis // *Journal of the American Oil Chemists Society*. 1987. 64(5): 630.

Forrest, 2015 – *Forrest M.D.* Why cancer cells have a more hyperpolarised mitochondrial membrane potential and emergent prospects for therapy // *BioRxiv*. 2015. 025197. DOI: <https://doi.org/10.1101/025197>

Franceschi et al., 2013 – *Franceschi P., Giordan M., Wehrens R.* Multiple comparisons in mass-spectrometry-based-omics technologies // *TrAC Trends in Analytical Chemistry*. 2013. 50: 11-21. DOI: <https://doi.org/10.1016/j.trac.2013.04.011>

Frey-Wyssling, 1965 – *Frey-Wyssling A.* Comparative organellography // *Experientia*. 1965. 21(12): 681-687. DOI: <https://doi.org/10.1007/bf02138470>

Friday et al., 2011 – *Friday E., Oliver III R., Welbourne T., Turturro F.* Glutaminolysis and glycolysis regulation by troglitazone in breast cancer cells: relationship to mitochondrial membrane potential. *Journal of cellular physiology*. 226(2): 511-519. DOI: <https://doi.org/10.1002/jcp.22360>

Gąbka et al., 2021 – *Gąbka M., Dalek P., Przybyło M., Gackowski D., Oliński R., Langner M.* The membrane electrical potential and intracellular ph as factors influencing intracellular ascorbate concentration and their role in cancer treatment // *Cells*. 2021. 10(11): 2964. DOI: <https://doi.org/10.3390/cells10112964>

Gayán et al., 2022 – *Gayán S., Joshi G., Dey T.* Biomarkers of mitochondrial origin: a futuristic cancer diagnostic // *Integrative Biology*. 2022. 14(4): 77-88. DOI: <https://doi.org/10.1093/intbio/zyac008>

Gekenidis et al., 2014 – *Gekenidis M.T., Studer P., Wüthrich S., Brunisholz R., Drissner D.* (2014). Beyond the matrix-assisted laser desorption ionization (MALDI) biotyping workflow: in search of microorganism-specific tryptic peptides enabling discrimination of subspecies // *Applied and environmental microbiology*. 2014. 80(14): 4234-4241. DOI: <https://doi.org/10.1128/aem.00740-14>

Girolamo et al., 2013 – *Girolamo F.D., Lante I., Muraca M., Putignani L.* The role of mass spectrometry in the “omics” era // *Current organic chemistry*. 2013. 17(23): 2891-2905. DOI: <https://doi.org/10.2174/1385272817888131118162725>

Gobert et al., 2021 – *Gobert A.P., Boutaud O., Asim M., Zagol-Ikapitte I.A., Delgado A.G., Latour Y.L., Finley J.L., Singh K., Verriere T.G., Allaman M.M., Barry, D.P.* Dicarbonyl electrophiles mediate inflammation-induced gastrointestinal carcinogenesis // *Gastroenterology*. 2021. 160(4): 1256-1268. DOI: <https://doi.org/10.1053/j.gastro.2020.11.006>

Gogichadze et al., 2014 – *Gogichadze G., Gogichadze T., Misabishvili E., Kamkamidze G.* (2014). Possible effect of variable membrane potential of a cancer cell on different carcinogenic processes // *Georgian medical news*. 204. 234: 116-120.

Gokulan et al., 2020 – *Gokulan R.C., Paulrasu K., Zaika E., Palrasu M., Boutaud O., Dikalov S., Zaika, A.* (2020). SU1171 - NADPH oxidases and reactive oxygen species generates isolog protein adducts in the esophagus: a novel mechanism that triggers esophageal carcinogenesis // *Gastroenterology*. 2020. 158(6): S-531.

Gradov, 2013 – *Gradov O.V.* Experimental Setups for Ozonometric Microscopy // *Biomedical Engineering*. 2013. 46(6): 260-264. DOI: <http://dx.doi.org/10.1007/s10527-013-9319-8>

Grigorov, 2012 – Grigorov B. Reactive oxygen species and their relation to carcinogenesis // *Trakia journal of sciences*. 2012. 10(3): 83-92.

Guerreiro et al., 2024 – Guerreiro E.M., Kruglik S.G., Swamy S., Latysheva N., Østerud B., Guigner J.M., Sureau F., Bonneau S., Kuzmin A.N., Prasad P.N., Hansen J.B. Extracellular vesicles from activated platelets possess a phospholipid-rich biomolecular profile and enhance prothrombinase activity // *Journal of Thrombosis and Haemostasis*. 2024. 22(5): 1463-1474. DOI: <https://doi.org/10.1016/j.jtha.2024.01.004>

Hadacek, Bachmann, 2015 – Hadacek F., Bachmann G. (2015). Low-molecular-weight metabolite systems chemistry // *Frontiers in environmental science*. 2015. 3: 12. DOI: <https://doi.org/10.3389/fenvs.2015.00012>

Harach et al., 2019 – Harach J.L., Lynch C.J., Montovano G., Olsen S.H., Foley T.D. Glyceraldehyde-3-phosphate dehydrogenase differentiates the cytotoxic mechanisms of cancer-active electrophiles in a yeast model // *The FASEB Journal*. 2019. 33(S1): 652-628. DOI: http://dx.doi.org/10.1096/fasebj.2019.33.1_supplement.652.8

Hong et al., 2021 – Hong J.K., Kim S.B., Lyou E.S., Lee T.K. Microbial phenomics linking the phenotype to function: The potential of Raman spectroscopy // *Journal of Microbiology*. 2021. 59: 249-258. DOI: <https://doi.org/10.1007/s12275-021-0590-1>

Houdelet, 2015 – Houdelet C., Bocquet M., Bulet P. MALDI Biotyping, an approach for deciphering and assessing the identity of the honeybee pathogen Nosema. *Rapid Communications in Mass Spectrometry*. 2015. 35(3): e8980. DOI: <http://dx.doi.org/10.1002/rcm.8980>

Huang, Spiers, 2006 – Huang W.E., Spiers A.J. Consideration of future requirements for Raman microbiology as an exemplar for the ab initio development of informatics frameworks for emergent OMICS technologies // *OMICS: A Journal of Integrative Biology*. 2006. 10(2): 238-241. DOI: <https://doi.org/10.1089/omi.2006.10.238>

Huang et al., 1994 – Huang X., Zhuang Z., Frenkel K., Klein C. B., Costa M. The role of nickel and nickel-mediated reactive oxygen species in the mechanism of nickel carcinogenesis // *Environmental health perspectives*. 1994. 102(Suppl 3): 281-284. DOI: <https://doi.org/10.1289/ehp.94102s3281>

Huang et al., 2023 – Huang L., Sun H., Sun L., Shi K., Chen Y., Ren X., Ge Y., Jiang D., Liu X., Knoll W., Zhang Q. (2023). Rapid, label-free histopathological diagnosis of liver cancer based on Raman spectroscopy and deep learning. *Nature Communications*. 2023. 14(1): 48. DOI: <https://doi.org/10.1038/s41467-022-35696-2>

Jablokov, Gradov, 2016 – Gradov O.V., Jablokow A.G. Novel morphometrics-on-a-chip: CCD- or CMOS-lab-on-a-chip based on discrete converters of different physical and chemical parameters of histological samples into the optical signals with positional sensitivity for morphometry of non-optical patterns // *Journal of Biomedical Technologies*. 2016. 2: 1-29. DOI: <http://dx.doi.org/10.15393/j6.art.2016.3642>

Jablokov, Gradov, 2015a – Jablokow A.G., Gradov O.V. MS-FRAP or MALDI Imaging Setups With Programmable Laser Sources: a New Way to the Diffusion, Molecular Mobility and Binding Measurements. In: Progr. 63-rd ASMS Conference on Mass Spectrometry and Allied Topics (St. Louis; June 2015), Section: "Imaging MS: Instrumentation" (JASMS Special Issue). 2015. DOI: <10.13140/RG.2.1.4919.1841>.

Jablokov, Gradov, 2015b – Jablokow A., Gradov O. Verifying Continuity of Membranous Organelles and Measurements of Exchange Rate Between the Nucleus and Cytoplasm using FLIP-Like MALDI-Based Imaging. In: Progr. 63-rd ASMS Conference on Mass Spectrometry and Allied Topics (St. Louis; June 2015), Section: "Imaging MS: Instrumentation" (JASMS Special Issue). 2015. DOI: <10.13140/RG.2.1.2322.3203>

Jablokov et al., 2017 – Jablokow A.G., Skrynnik A.A., Orekhov F.K., Nasirov P.A., Gradov O.V. "MALDI-FLIP-on-a-chip" and " MALDI-FRAP-on-a-flap": Novel Techniques for Soil Microbiology and Environmental Biogeochemistry. I-MALDI Chip Fingerprinting. *Biogeosystem Technique*. 2017. 4: 140-188. DOI: <https://doi.org/10.13187/bgt.2017.2.140>

Jablokov et al., 2018 – Jablokow A.G., Nasirov P.A., Orekhov F.K., Gradov O.V. "MALDI-FLIP-on-a-chip" and " MALDI-FRAP-on-a-flap": Novel Techniques for Soil Microbiology and Environmental Biogeochemistry. II-Polymer Chip Prototyping // *Biogeosystem Technique*. 2018. (5): 3-56. DOI: <https://doi.org/10.13187/bgt.2018.1.3>

Jaeken, 2017 – Jaeken L. The neglected functions of intrinsically disordered proteins and the origin of life // *Progress in Biophysics and Molecular Biology*. 2017. 126: 31-46. DOI: <https://doi.org/10.1016/j.pbiomolbio.2017.03.002>

Janků et al., 2019a – Janků M., Luhová L., Petřivalský M. On the origin and fate of reactive oxygen species in plant cell compartments. *Antioxidants*. 2019. 8(4): 105. DOI: <https://doi.org/10.3390/antiox8040105>

Janků et al., 2019b – Janků M., Tichá T., Luhová L., Petřivalský M. Chapter 40: Compartmentalization of reactive Oxygen species and Nitric Oxide production in plant cells // *Reactive oxygen, nitrogen and sulfur species in plants: production, Metabolism, Signaling and Defense Mechanisms*. 2019: 923-945. DOI: <https://doi.org/10.1002/9781119468677.ch40>

Jian, 2023 – Jian X.U. Ramanome, FlowRACS and RACS-Seq/Culture: functional dissection and mining of microbiomes at single-cell resolution. 2023. [Electronic resource]. URL: https://www.ramanfestconf.com/2023/Abstracts/2023_Xu.pdf

Jones, Sies, 2015 – Jones D.P., Sies H. The redox code // *Antioxidants & redox signaling*. 2015. 23(9): 734-746. DOI: <https://doi.org/10.1089/ars.2015.6247>

Jopkiewicz, 2019 – Jopkiewicz S.P. Significance of reactive oxygen species and oxidative stress in carcinogenesis // *Journal of Education, Health and Sport*. 2019. 9(6): 241-250. DOI: <http://dx.doi.org/10.5281/zenodo.3243595>

Klein, Costa, 1991 – Klein C.B., Costa M. The Role of Nickel and Nickel-mediated Reactive Oxygen Species in the Mechanism of Nickel Carcinogenesis // *Environmental Health Perspectives*. 1991. 102: 281. DOI: <https://doi.org/10.1289/ehp.94102s3281>

Kosmachevskaya, Topunov, 2021 – Kosmachevskaya O.V., Topunov A.F. Nonenzymatic reactions in metabolism: Their role in evolution and adaptation // *Applied Biochemistry and Microbiology*. 2021. 57: 543-555. DOI: <http://dx.doi.org/10.1134/S0003683821050100>

Kovacic, Abadjian, 2018 – Kovacic P., Abadjian M.C.Z. Mechanism of lung carcinogenesis: electron transfer, reactive oxygen species, oxidative stress and antioxidants // *SOJ Microbiol Infect Dis*. 2018. 2: 1-10.

Kovacic et al., 1986 – Kovacic P., Crawford P.W., Ryan M.D., Nelson V.C. 844—Charge transfer mechanism for carcinogenesis by alkylating and other agents // *Bioelectrochemistry and Bioenergetics*. 1986. 15(3): 305-316. DOI: [https://doi.org/10.1016/0302-4598\(86\)85020-6](https://doi.org/10.1016/0302-4598(86)85020-6)

Kruk, Aboul-Enein, 2017 – Kruk J., Aboul-Enein H. Reactive oxygen and nitrogen species in carcinogenesis: implications of oxidative stress on the progression and development of several cancer types // *Mini reviews in medicinal chemistry*. 2017. 17(11): 904-919. DOI: <https://doi.org/10.2174/138955751766170228115324>

Kuwahara et al., 2021 – Kuwahara Y., Tomita K., Roudkenar M.H., Roushandeh A.M., Urushihara Y., Igarashi K., Kurimasa A., Sato T. Decreased mitochondrial membrane potential is an indicator of radioresistant cancer cells // *Life Sciences*. 2021. 286: 120051. DOI: <https://doi.org/10.1016/j.lfs.2021.120051>

Kuzmin et al., 2017a – Kuzmin A.N., Pliss A., Prasad P.N. Ramanomics: new omics disciplines using micro Raman spectrometry with biomolecular component analysis for molecular profiling of biological structures // *Biosensors*. 2017. 7(4): 52. DOI: <https://doi.org/10.3390/bios7040052>

Kuzmin et al., 2017b – Kuzmin A.N., Levchenko S.M., Pliss A., Qu J., Prasad P.N. Molecular profiling of single organelles for quantitative analysis of cellular heterogeneity // *Scientific Reports*. 2017. 7(1): 6512. DOI: <https://doi.org/10.1038/s41598-017-06936-z>

Kuzmin et al., 2018 – Kuzmin A.N., Pliss A., Rzhevskii A., Lita A., Larion M. BCAbx algorithm expands capabilities of Raman microscope for single organelles assessment // *Biosensors*. 2018. 8(4): 106. DOI: <https://doi.org/10.3390/bios8040106>

LaLone et al., 2023 – LaLone V., Aizenshtadt A., Goertz J., Skottvoll F.S., Mota M.B., You J., Zhao X., Berg H.E., Stokowiec J., Yu M., Schwendeman A. (2023). Quantitative chemometric phenotyping of three-dimensional liver organoids by Raman spectral imaging // *Cell Reports Methods*. 2023. 3(4): 100440. DOI: <https://doi.org/10.1016/j.crmeth.2023.100440>

LaLone et al., 2019a – LaLone V., Fawaz M.V., Morales-Mercado J., Mourão M.A., Snyder C.S., Kim S.Y., Lieberman A.P., Tuteja A., Mehta G., Standiford T.J., Raghavendran K. Inkjet-printed micro-calibration standards for ultraquantitative Raman spectral cytometry. *Analyst*. 2019. 144(12): 3790-3799. DOI: <https://doi.org/10.1039/c9an00500e>

LaLone et al., 2019b – LaLone V., Mourão M.A., Standiford T.J., Raghavendran K., Shedd K., Stringer K.A., Rosania G.R. An expandable mechanopharmaceutical device (3): A versatile Raman spectral cytometry approach to study the drug cargo capacity of individual macrophages // *Pharmaceutical Research*. 2019. 36(1): 2. DOI: <https://doi.org/10.1007/s11095-018-2540-0>

Lareau, 2016 – Lareau N.M. Development of Ion Mobility and Mass Spectrometry Strategies in Support of Integrated Omics and Systems Biology. Vanderbilt University, 2016.

Larion et al., 2018 – Larion M., Dowdy T., Ruiz-Rodado V., Meyer M.W., Song H., Zhang W., Davis D., Gilbert M.R., Lita A. Detection of metabolic changes induced via drug treatments in live cancer cells and tissue using Raman imaging microscopy // *Biosensors*. 2018. 9(1): 5. DOI: <https://doi.org/10.3390/bios9010005>

Lawrence, 2023 – Lawrence C.P. Simple machine learning methods work surprisingly well for Ramanomics // *Journal of Raman Spectroscopy*. 2023. 54(8): 887-889. DOI: <https://doi.org/10.1002/jrs.6555>

Lee et al., 2016 – Lee D.G., Choi B.K., Kim Y.H., Oh H.S., Park S.H., Bae Y.S., Kwon B.S. The repopulating cancer cells in melanoma are characterized by increased mitochondrial membrane potential // *Cancer letters*. 2016. 382(2): 186-194. DOI: <https://doi.org/10.1016/j.canlet.2016.08.027>

Lei et al., 2021 – Lei L., Zhang J., Decker E.A., Zhang G. Roles of lipid peroxidation-derived electrophiles in pathogenesis of colonic inflammation and colon cancer // *Frontiers in Cell and Developmental Biology*. 2021. 9: 665591. DOI: <https://doi.org/10.3389/fcell.2021.665591>

Lemeshko, 2015 – Lemeshko V.V. Channeling of mitochondrial energy in cardiac and cancer cells by the metabolically-dependent outer membrane potential // *Biophysical Journal*. 2015. 108(2): 607a.

Lennicke et al., 2016 – Lennicke C., Rahn J., Heimer N., Lichtenfels R., Wessjohann L.A., Seliger B. Redox proteomics: Methods for the identification and enrichment of redox-modified proteins and their applications // *Proteomics*. 2016. 16(2): 197-213. DOI: <https://doi.org/10.1002/pmic.201500268>

Li et al., 2020 – Li X., Yang F., Rubinsky B. A correlation between electric fields that target the cell membrane potential and dividing HeLa cancer cell growth inhibition // *IEEE Transactions on Biomedical Engineering*. 2020. 68(6): 1951-1956. DOI: <https://doi.org/10.1109/tbme.2020.3042650>

Lorenz et al., 2020 – Lorenz B., Ali N., Bocklitz T., Rösch P., Popp J. Discrimination between pathogenic and non-pathogenic *E. coli* strains by means of Raman microspectroscopy // *Analytical and Bioanalytical Chemistry*. 2020. 412: 8241-8247. DOI: <https://doi.org/10.1007/s00216-020-02957-2>

Lu et al., 2020 – Lu W., Chen X., Wang L., Li H., Fu Y.V. Combination of an artificial intelligence approach and laser tweezers Raman spectroscopy for microbial identification // *Analytical chemistry*. 2020. 92(9): 6288-6296. DOI: <https://doi.org/10.1021/acs.analchem.9b04946>

Lu et al., 2023 – Lu H., Zhang H., Li L. Chemical tagging mass spectrometry: an approach for single-cell omics // *Analytical and bioanalytical chemistry*. 2023. 415(28): 6901-6913. DOI: <https://doi.org/10.1007/s00216-023-04850-0>

Luo et al., 2022 – Luo Y., Sobhani Z., Zhang Z., Zhang X., Gibson C.T., Naidu R., Fang C. Raman imaging and MALDI-MS towards identification of microplastics generated when using stationery markers // *Journal of Hazardous Materials*. 2022. 424: 127478. DOI: <https://doi.org/10.1016/j.jhazmat.2021.127478>

Lyublinskaya, Antunes, 2019 – Lyublinskaya O., Antunes F. Measuring intracellular concentration of hydrogen peroxide with the use of genetically encoded H₂O₂ biosensor HyPer // *Redox biology*. 2019. 24: 101200. DOI: <https://doi.org/10.1016/j.redox.2019.101200>

MacLeod et al., 2009 – MacLeod A.K., McMahon M., Plummer S.M., Higgins L.G., Penning T.M., Igarashi K., Hayes J.D. Characterization of the cancer chemopreventive NRF2-dependent gene battery in human keratinocytes: demonstration that the KEAP1–NRF2 pathway, and not the BACH1–NRF2 pathway, controls cytoprotection against electrophiles as well as redox-cycling compounds // *Carcinogenesis*. 2009. 30(9): 1571-1580. DOI: <https://doi.org/10.1093/carcin/bgp176>

Mallah et al., 2023 – Mallah K., Zibara K., Kerbaj C., Eid A., Khoshman N., Ousseily Z., Kobeissy A., Cardon T., Cizkova D., Kobeissy F., Fournier I. Neurotrauma investigation through spatial omics guided by mass spectrometry imaging: Target identification and clinical applications // *Mass spectrometry reviews*. 2023. 42(1): 189-205. DOI: <https://doi.org/10.1002/mas.21719>

Marino et al., 1994 – *Marino A.A., Iliev I.G., Schwalke M.A., Gonzalez E., Marler K.C., Flanagan C.A.* Association between cell membrane potential and breast cancer // *Tumor Biology*. 1994. 15(2): 82-89. DOI: <https://doi.org/10.1159/000217878>

Marquez-Quinones, 2007 – *Marquez-Quinones A.* Reactive oxygen species, hepatitis and carcinogenesis initiation: an integrative approach combining transcriptomic and metabonomic profilings (Doctoral dissertation, Institut National des Sciences Appliquées de Toulouse). 2007. [Electronic resource]. URL: <https://theses.fr/2007ISAT0028>

Medeiros, 2018 – *Medeiros F.H.C.* The role of reactive oxygen species in thyroid radio-carcinogenesis (Doctoral dissertation, Université Paris-Saclay (ComUE); Universidade federal do Rio de Janeiro). 2018. [Electronic resource]. URL: <https://theses.fr/2018SACLS085; https://theses.hal.science/tel-04010752> (NNT : 2018SACLS085).

Miller, 1998 – *Miller J.A.* The metabolism of xenobiotics to reactive electrophiles in chemical carcinogenesis and mutagenesis: a collaboration with Elizabeth Cavert Miller and our associates // *Drug metabolism reviews*. 1998. 30(4): 645-674. DOI: <https://doi.org/10.3109/03602539808996326>

Moldogazieva et al., 2018 – *Moldogazieva N.T., Lutsenko S.V., Terentiev A.A.* Reactive oxygen and nitrogen species-induced protein modifications: implication in carcinogenesis and anticancer therapy // *Cancer Research*. 2018. 78(21): 6040-6047. DOI: <https://doi.org/10.1158/0008-5472.can-18-0980>

Mosier-Boss, 2017 – *Mosier-Boss P.A.* Review on SERS of Bacteria // *Biosensors*. 2017. 7(4): 51. DOI: <https://doi.org/10.3390/bios7040051>

Murray, Van Eyk, 2012 – *Murray C.I., Van Eyk J.E.* Chasing cysteine oxidative modifications: proteomic tools for characterizing cysteine redox status // *Circulation: Cardiovascular Genetics*. 2012. 5(5): 591. DOI: <https://doi.org/10.1161/CIRCGENETICS.111.961425>

Nie et al., 2016 – *Nie W., Yan L., Lee Y.H., Guha C., Kurland I.J., Lu H.* Advanced mass spectrometry-based multi-omics technologies for exploring the pathogenesis of hepatocellular carcinoma // *Mass Spectrometry Reviews*. 2016. 35(3): 331-349. DOI: <https://doi.org/10.1002/mas.21439>

Nishigori et al., 2004 – *Nishigori C., Hattori Y., Toyokuni S.* Role of reactive oxygen species in skin carcinogenesis // *Antioxidants and Redox Signaling*. 2004. 6(3): 561-570. DOI: <https://doi.org/10.1089/152308604773934314>

Nnodim, Hauwa, 2020 – *Nnodim J., Hauwa B.* Membrane potential: An emerging and important player in cancer metastasis // *Asclep Med Res Rev*. 2020. 3: 1-2.

Okada, 2007 – *Okada F.* Beyond foreign-body-induced carcinogenesis: impact of reactive oxygen species derived from inflammatory cells in tumorigenic conversion and tumor progression // *International journal of cancer*. 2007. 121(11) 2364-2372. DOI: <https://doi.org/10.1002/ijc.23125>

Okazaki, 2022 – *Okazaki Y.* Asbestos-induced mesothelial injury and carcinogenesis: Involvement of iron and reactive oxygen species // *Pathology International*. 2022. 72(2): 83-95. DOI: <https://doi.org/10.1111/pin.13196>

Oliński, Jurgowiak, 1999 – *Oliński R., Jurgowiak M.* The role of reactive oxygen species in mutagenesis and carcinogenesis processes // *Postepy Biochemii*. 1999. 45(1): 50-58.

Olsen et al., 2018 – *Olsen S., Foley T., Montovano G., Lynch C.* Use of phenylarsine oxide-affinity chromatography to identify common cellular targets of cancer-active electrophiles // *Free Radical Biology and Medicine*. 2018. 128: S72. DOI: <https://doi.org/10.1016/j.freeradbiomed.2018.10.154>

Orekhov, Gradov, 2022 – *Orekhov F., Gradov O.* Automated Soil Microbiology Using Lensless and LDI MS Imaging with Buried Slides // *Smart Innovation, Systems and Technologies*. 2022. 247: 471-479. DOI: http://dx.doi.org/10.1007/978-981-16-3844-2_43

Orekhov, Gradov, 2023 – *Orekhov F.K., Gradov O.V.* Towards Ultraviolet Microbeam Scanning and Lens-Less UV Microbeam Microscopy with Mirror Galvanometric Scanners: From the History of Research Instrumentation to Engineering of Modern Mechatronic Optical Systems. // *J Sen Net Data Comm*. 2023. 3(1): 117-137.

Orekhov, Gradov, 2023a – *Orekhov F.K., Gradov O.V.* Target Chip Based Single-Cell Biotyping and Telemetric Bioluminescence Lensless Microscopy of the Buried Sandwich-Slides as a Novel Way for Measurement, Mapping and Molecular Imaging of Biodegradation / Biofouling of Plastic Surfaces in Real Soils // *Advances in Transdisciplinary Engineering*. 2023. 38: 417-425. DOI: <http://dx.doi.org/10.3233/ATDE230317>

[Orekhov, Gradov, 2023b](#) – Orekhov T.K., Gradov O.V. From desolvation-induced self-organization on the MALDI anchor target chip surfaces to laser-induced self-organization in MALDI techniques: Correlation-spectral analysis and complex wavelet analysis of tesiographic spots on the anchor chips // *Materials Technology Reports*. 2023. 1(1): 124. DOI: <http://dx.doi.org/10.59400/mtr.vii.124>

[Orekhov et al., 2016](#) – Orekhov F.K., Jablokow A.G., Skrynnik A.A. Hybridization of laser-induced spectrofluorescence analysis (LIFS), matrix-assisted laser desorption/ionization mass spectrometry (MALDI), fluorescence recovery after photobleaching (FRAP) and fluorescence loss in photobleaching (FLIP) microtechnics // *Journal of Biomedical Technologies*. 2016. 2: 42-52. DOI: <http://dx.doi.org/10.15393/j6.art.2016.3702>

[Orekhov et al., 2023](#) – Orekhov F.K., Jablokow A.G., Gradow O.V. "Novel MALDI MS + FLIP approaches for verifying continuity of membranous structures and measurements of nucleus-cytoplasm exchange" (Poster). At: IMSIS 2023 (October 23-25, Montreal). Program and Abstracts. 2023. DOI: [10.13140/RG.2.2.31971.53287](https://doi.org/10.13140/RG.2.2.31971.53287)

[Pade et al., 2021](#) – Pade L.R., Stepler K.E., Portero E.P., DeLaney K., Nemes P. Biological mass spectrometry enables spatiotemporal ‘omics: From tissues to cells to organelles // *Mass spectrometry reviews*. 2021. 43(1): 106-138. DOI: <https://doi.org/10.1002/mas.21824>

[Paglia et al., 2022](#) – Paglia G., Smith A.J., Astarita G. Ion mobility mass spectrometry in the omics era: Challenges and opportunities for metabolomics and lipidomics // *Mass Spectrometry Reviews*. 2022. 41(5): 722-765. DOI: <https://doi.org/10.1002/mas.21686>

[Panayiotidis, 2008](#) – Panayiotidis M.I. Reactive oxygen species (ROS) in multistage carcinogenesis // *Cancer letters*. 2008. 266(1): 3-5. DOI: <https://doi.org/10.1016/j.canlet.2008.02.027>

[Paulech et al., 2013](#) – Paulech J., Solis N., Edwards A.V., Puckeridge M., White M.Y., Cordwell S.J. Large-scale capture of peptides containing reversibly oxidized cysteines by thiol-disulfide exchange applied to the myocardial redox proteome // *Analytical chemistry*. 2013. 85(7): 3774-3780. DOI: <https://doi.org/10.1021/ac400166e>

[Pena et al., 2022](#) – Pena I., Pena-Vina E., Rodriguez-Avial I., Picazo J.J., Gómez-González Á., Culebras E. Comparison of performance of MALDI-TOF MS and MLST for biotyping carbapenemase-producing Klebsiella pneumoniae sequence types ST11 and ST101 isolates. *Enfermedades infecciosas y microbiologia clinica (English ed.)*. 2022. 40(4): 172-178. DOI: <https://doi.org/10.1016/j.eimce.2020.10.011>

[Person et al., 2003](#) – Person M.D., Monks T.J., Lau S.S. An integrated approach to identifying chemically induced posttranslational modifications using comparative MALDI-MS and targeted HPLC-ESI-MS/MS // *Chemical research in toxicology*. 2003. 16(5): 598-608. DOI: <https://doi.org/10.1021/tx020109f>

[Phulara, Seneviratne, 2024](#) – Phulara N.R., Seneviratne H.K. Mass spectrometry imaging-based multi-omics approaches to understand drug metabolism and disposition // *Journal of Mass Spectrometry*. 2024. 59(7): e5042. DOI: <https://doi.org/10.1002/jms.5042>

[Pliss et al., 2021](#) – Pliss A., Kuzmin A.N., Lita A., Kumar R., Celiku O., Atilla-Gokcumen G.E., Gokcumen O., Chandra D., Larion M., Prasad P.N. A single-organelle optical omics platform for cell science and biomarker discovery. *Analytical chemistry*. 2021. 93(23): 8281-8290. DOI: <https://doi.org/10.1021/acs.analchem.1c01131>

[Portugalov et al., 1964](#) – Portugalov V.V., Krasnov I.B., Ball' T.V. A histochemical determination of the denaturation of the proteins of the nerve cell induced by alcohol // *Bulletin of Experimental Biology and Medicine*. 1964. 55(4): 458-460. DOI: <https://doi.org/10.1007/BF00785679>

[Pralea et al., 2020](#) – Pralea I.E., Moldovan R.C., Tigu A.B., Ionescu C., Iuga C.A. Mass spectrometry-based omics for the characterization of triple-negative breast cancer bio-signature // *Journal of personalized medicine*. 2020. 10(4): 277. DOI: <https://doi.org/10.3390/jpm10040277>

[Pranada et al., 2016](#) – Pranada A.B., Schwarz G., Kostrzewska M. MALDI Biotyping for microorganism identification in clinical microbiology // *Advances in MALDI and laser-induced soft ionization mass spectrometry*. 2016. 197-225. DOI: [10.1007/978-3-319-04819-2_11](https://doi.org/10.1007/978-3-319-04819-2_11)

[Quanico et al., 2017](#) – Quanico J., Franck J., Wisztorski M., Salzet M., Fournier I. (2017). Integrated mass spectrometry imaging and omics workflows on the same tissue section using grid-aided, parafilm-assisted microdissection // *Biochimica et Biophysica Acta (BBA)-General Subjects*. 2017. 1861(7): 1702-1714. DOI: <https://doi.org/10.1016/j.bbagen.2017.03.006>

Quintá et al., 2016 – Quintá H.R., Wilson C., Blidner A.G., González-Billault C., Pasquini L.A., Rabinovich G.A., Pasquini J.M. Ligand-mediated Galectin-1 endocytosis prevents intraneuronal H₂O₂ production promoting F-actin dynamics reactivation and axonal re-growth // *Experimental neurology*. 2016. 283: 165-178. DOI: <https://doi.org/10.1016/j.expneurol.2016.06.009>

Ralph et al., 2010 – Ralph S.J., Rodríguez-Enríquez S., Neuzil J., Saavedra E., Moreno-Sánchez R. The causes of cancer revisited: “mitochondrial malignancy” and ROS-induced oncogenic transformation—why mitochondria are targets for cancer therapy // *Molecular aspects of medicine*. 2010. 31(2): 145-170. DOI: <https://doi.org/10.1016/j.mam.2010.02.008>

Rezende et al., 2018 – Rezende F., Brandes R.P., Schröder K. Detection of hydrogen peroxide with fluorescent dyes // *Antioxidants & Redox Signaling*. 2018. 29(6): 585-602. DOI: <https://doi.org/10.1089/ars.2017.7401>

Rhee et al., 2010 – Rhee S.G., Chang T.S., Jeong W., Kang D. Methods for detection and measurement of hydrogen peroxide inside and outside of cells // *Molecules and cells*. 2010. 29: 539-549. DOI: <https://doi.org/10.1007/s10059-010-0082-3>

Ryabchikov et al., 2018 – Ryabchikov O., Popp J., Bocklitz T. Fusion of MALDI spectrometric imaging and Raman spectroscopic data for the analysis of biological samples // *Frontiers in Chemistry*. 2018. 6: 257. DOI: <https://doi.org/10.3389/fchem.2018.00257>

Sadri et al., 2022 – Sadri H., Aghaei M., Akbari V. Nisin induces apoptosis in cervical cancer cells via reactive oxygen species generation and mitochondrial membrane potential changes // *Biochemistry and Cell Biology*. 2022. 100(2): 136-141. DOI: <https://doi.org/10.1139/bcb-2021-0225>

Samoylenko et al., 2013 – Samoylenko A., Hossain J.A., Mennerich D., Kellokumpu S., Hiltunen J. K., Kietzmann T. Nutritional countermeasures targeting reactive oxygen species in cancer: from mechanisms to biomarkers and clinical evidence // *Antioxidants & redox signaling*. 2013. 19(17): 2157-2196. DOI: <https://doi.org/10.1089/ars.2012.4662>

Sanders, Edwards, 2020 – Sanders K.L., Edwards J.L. Nano-liquid chromatography-mass spectrometry and recent applications in omics investigations // *Analytical Methods*. 2020. 12(36): 4404-4417. DOI: <https://doi.org/10.1039/doay01194k>

Schmitt et al., 2023 – Schmitt R., Qayum S., Pliss A., Kuzmin A.N., Muthiah V.P.K., Kaliyappan K., Prasad P.N., Mahajan S.D. Mitochondrial Dysfunction and Apoptosis in Brain Microvascular Endothelial Cells Following Blast Traumatic Brain Injury // *Cellular and Molecular Neurobiology*. 2023. 43(7): 3639-3651. DOI: <https://doi.org/10.1007/s10571-023-01372-2>

Schulte-Hermann et al., 2006 – Schulte-Hermann R., Teufelhofer O., Parzefall W., Freiler C., Grasl-Kraupp B., Gerner C. (2006). Reactive oxygen from macrophages and chemical carcinogenesis // *Toxicology Letters*. 2006. (164): S4. DOI: <http://dx.doi.org/10.1016/j.toxlet.2006.06.012>

Shen et al., 2021 – Shen Y., Yue J., Xu W., Xu S. (2021). Recent progress of surface-enhanced Raman spectroscopy for subcellular compartment analysis. *Theranostics*. 11(10): 4872. DOI: <https://doi.org/10.7150/thno.56409>

Shi et al., 1998 – Shi X., Castranova V., Halliwell B., Valliyathan V. Reactive oxygen species and silica-induced carcinogenesis *Journal of Toxicology and Environmental Health, Part B Critical Reviews*. 1(3): 181-197. DOI: <https://doi.org/10.1080/10937409809524551>

Shimura, Ushiyama, 2024 – Shimura T., Ushiyama A. Mitochondrial reactive oxygen species-mediated fibroblast activation has a role in tumor microenvironment formation in radiation carcinogenesis. *Radiation Protection Dosimetry*. 2024. 200(16-18): 1590-1593. DOI: <https://doi.org/10.1093/rpd/ncae027>

Shkarina et al., 1984 – Shkarina T.N., Zatsepina G.N., Kasatkina V.V., Kozinets G.I., Tarasova I.M. (1984). Electrical charge of the surface of lymphocytes and their capacity for mitotic division in induced carcinogenesis. *Biophysics*. 1984. 29(1): 102-106.

Siddhanta et al., 2023 – Siddhanta S., Kuzmin A.N., Pliss A., Baev A.S., Khare S.K., Chowdhury P.K., Ganguli A.K., Prasad P.N. (2023). Advances in Raman spectroscopy and imaging for biomedical research. *Advances in Optics and Photonics*. 2023. 15(2): 318-384. DOI: <http://dx.doi.org/10.1364/AOP.479884>

Skates, 2022 – Skates E. Measurement and manipulation of mitochondrial membrane potential in cancer cells (Doctoral dissertation, University of Warwick), 2022.

Skottvoll, 2022 – Skottvoll F.S. Liver organoids, mass spectrometry, and separation science (Dissertation for the degree Philosophiae Doctor). University of Oslo (Department of Chemistry; Faculty of Mathematics and Natural Sciences). DOI: <https://doi.org/10.1002/anse.202100051>

- Smets et al., 2021** – Smets T., De Keyser T., Tousseyn T., Waelkens E., De Moor B.. Correspondence-aware manifold learning for microscopic and spatial omics imaging: a novel data fusion method bringing mass spectrometry imaging to a cellular resolution // *Analytical Chemistry*. 2021. 93(7): 3452-3460. DOI: <https://doi.org/10.1021/acs.analchem.oco4759>
- Smith et al., 2014** – Smith M.R., Zhou F., Kumar P.V., Beggs R., Velu S., Landar A., Murphy M. (2014). Metabolic Reprogramming by a Mitochondria-Targeted Electrophile in Breast Cancer Cells // *Free Radical Biology and Medicine*. 2014. (76): S132. DOI: <http://dx.doi.org/10.1016/j.freeradbiomed.2014.10.212>
- Smolyarova et al., 2022** – Smolyarova D.D., Podgorny O.V., Bilan D.S., Belousov V.V. (2022). A guide to genetically encoded tools for the study of H₂O₂ // *The FEBS journal*. 2022. 289(18): 5382-5395. DOI: <https://doi.org/10.1111/febs.16088>
- Somboro et al., 2014** – Somboro A.M., Essack S.Y., Tiwari D., Shobo A., Bester L.A., Kruger H.G., Govender T. (2014). Evaluation of MALDI Biotyping for Rapid Subspecies Identification of Carbapenemase-Producing Bacteria via Protein Profiling // *Mass Spectrometry Letters*. 2014. 5(4): 110-114. DOI: <http://dx.doi.org/10.5478/MSL.2014.5.4.110>
- Standeven, Wetterhahn, 1991** – Standeven A.M., Wetterhahn K.E. Is there a role for reactive oxygen species in the mechanism of chromium (VI) carcinogenesis? // *Chemical research in toxicology*. 1991. 4(6): 616-625. DOI: <https://doi.org/10.1021/tx00024a003>
- Tamura et al., 2013** – Tamura M., Mutoh M., Fujii G., Matsui H. Involvement of Mitochondrial Reactive Oxygen Species in Gastric Carcinogenesis // *J Gastroint Dig Syst*. 2013. 3: 150. DOI: <https://doi.org/10.4172/2161-069X.1000150>
- Tian et al., 2021** – Tian Y., Xu W., Ma K., Cong L., Shen Y., Han X., Liang C., Liang L., Qi G., Jin Y., Xu S. Label-free analysis of cell membrane proteins via evanescent field excited surface-enhanced Raman scattering // *The Journal of Physical Chemistry Letters*. 2021. 12(43): 10720-10727. DOI: <https://doi.org/10.1021/acs.jpclett.1c02966>
- Tokuoka, Morioka, 1957** – Tokuoka S., Morioka H. The membrane potential of the human cancer and related cells (I) // *Gann*. 1957. 48(4): 353-354.
- Valavanidis, 1994** – Valavanidis A. Ultraviolet radiation and skin cancer implication of free radical reactions and reactive oxygen species in skin carcinogenesis // *Review of Clinical Pharmacology and Pharmacokinetics (International Edition)*. 1994. 8: 101. DOI: <https://doi.org/10.1042/bss0610047>
- Valavanidis, 2019** – Valavanidis A. Oxidative stress and pulmonary carcinogenesis through mechanisms of reactive oxygen species. How respirable particulate matter, fibrous dusts, and ozone cause pulmonary inflammation and initiate lung carcinogenesis // *Oxidative Stress in Lung Diseases*. 2019. 1: 247-265. DOI: https://doi.org/10.1007/978-981-13-8413-4_13
- Vostrikova et al., 2020** – Vostrikova S.M., Grinev A.B., Gogvadze V.G. Reactive oxygen species and antioxidants in carcinogenesis and tumor therapy // *Biochemistry (Moscow)*. 2020. 85: 1254-1266. DOI: <https://doi.org/10.1134/s0006297920100132>
- Wang et al., 2023a** – Wang X., Han J., Li Z., Li B., Wan Y., Liu L. (2023). Insight into plant spatial omics: mass spectrometry imaging // *Frontiers in Plant Science*. 2023. 14: 1273010. DOI: <https://doi.org/10.3389/fpls.2023.1273010>
- Wang et al., 2022** – Wang J., Pursell M.E., DeVor A., Awoyemi O., Valentine S.J., Li P. (2022). Portable mass spectrometry system: instrumentation, applications, and path to ‘omics analysis // *Proteomics*. 2022. 22(23-24): 2200112. DOI: <https://doi.org/10.1002/pmic.202200112>
- Wang et al., 2023b** – Wang Z., Liu B., Lin L., Qiao L. Mass spectrometry for mitochondrial multi-omics. *TrAC Trends in Analytical Chemistry*. 163: 117063. DOI: <https://doi.org/10.1016/j.trac.2023.117063>
- Wang, 2009** – Wang G. NADPH oxidase and reactive oxygen species as signaling molecules in carcinogenesis. *Frontiers of Medicine in China*. 2009. 3: 1-7. DOI: <https://doi.org/10.1007/s11684-009-0018-5>
- Weller et al., 2014** – Weller J., Kizina K.M., Can K., Bao G., Müller M. Response properties of the genetically encoded optical H₂O₂ sensor HyPer // *Free Radical Biology and Medicine*. 2014. 76: 227-241. DOI: <https://doi.org/10.1016/j.freeradbiomed.2014.07.045>
- Wolyniak et al., 2018** – Wolyniak M.J., Reyna N.S., Plymale R., Pope W.H., Westholm D.E. Mass spectrometry as a tool to enhance “-omics” education // *Journal of Microbiology & Biology Education*. 2018. 19(1): 10-1128. DOI: <https://doi.org/10.1128/jmbe.v19i1.1459>

Wu, Ni, 2015 – Wu Q., Ni X. (2015). ROS-mediated DNA methylation pattern alterations in carcinogenesis. *Current drug targets*. 2015. 16(1): 13-19. DOI: <https://doi.org/10.2174/138945011666150113121054>

Yang, Brackenbury, 2013 – Yang M., Brackenbury W.J. Membrane potential and cancer progression // *Frontiers in physiology*. 2013. 4: 185. DOI: <https://doi.org/10.3389/fphys.2013.00185>

Yang et al., 2023 – Yang E., Kim J.H., Tressler C.M., Shen X.E., Brown D.R., Johnson C.C., Hahm T.H., Barman I., Glunde K. RaMALDI: enabling simultaneous Raman and MALDI imaging of the same tissue section // *Biosensors and Bioelectronics*. 2023. 239: 115597. DOI: <https://doi.org/10.1016/j.bios.2023.115597>

Yang, 2014 – Yang Y. Alternative Approaches to Optical Sensing of the Redox State. In *Natural Biomarkers for Cellular Metabolism*. 2014. Pp. 208-229. CRC Press. DOI: <http://dx.doi.org/10.1201/b17427-11>

Ye et al., 2011 – Ye X.Q., Wang G.H., Huang G.J., Bian X.W., Qian G.S., Yu S.C.). Heterogeneity of mitochondrial membrane potential: a novel tool to isolate and identify cancer stem cells from a tumor mass? // *Stem Cell Reviews and Reports*. 2011. 7: 153-160. DOI: <https://doi.org/10.1007/s12015-010-9122-9>

Zaikin, Borisov, 2021 – Zaikin V.G., Borisov R.S. Mass spectrometry as a crucial analytical basis for omics sciences // *Journal of Analytical Chemistry*. 2021. 76: 1567-1587. DOI: <https://doi.org/10.1134/S1061934821140094>

Zhang, Qiao, 2024 – Zhang D., Qiao L. Microfluidics coupled mass spectrometry for single cell multi-omics // *Small Methods*. 2024. 8(1): 2301179. DOI: <https://doi.org/10.1002/smtd.202301179>

Zhang et al., 2007 – Zhang X., Wei D., Yap Y., Li L., Guo S., Chen F. Mass spectrometry-based “omics” technologies in cancer diagnostics // *Mass spectrometry reviews*. 2007. 26(3): 403-431. DOI: <https://doi.org/10.1002/mas.20132>

Zhang et al., 2019 – Zhang P., Wang L., Fang Y., Zheng D., Lin T., Wang H. Label-free exosomal detection and classification in rapid discriminating different cancer types based on specific Raman phenotypes and multivariate statistical analysis // *Molecules*. 2019. 24(16): 2947. DOI: <https://doi.org/10.3390/molecules24162947>

Zhang et al., 2023 – Zhang H., Delafield D.G., Li L. Mass spectrometry imaging: the rise of spatially resolved single-cell omics // *Nature Methods*. 2023. 20(3): 327-330. DOI: <https://doi.org/10.1038/s41592-023-01774-6>

Zhao, Cai, 2023 – Zhao C., Cai Z. Mass spectrometry-based omics and imaging technique: a novel tool for molecular toxicology and health impacts // *Reviews of Environmental Contamination and Toxicology*. 2023. 261(1): 10. DOI: <http://dx.doi.org/10.1007/s44169-023-00032-2>

Zhao et al., 2022 – Zhao C., Dong J., Deng L., Tan Y., Jiang W., Cai Z. Molecular network strategy in multi-omics and mass spectrometry imaging // *Current Opinion in Chemical Biology*. 2022. 70: 102199. DOI: <https://doi.org/10.1016/j.cbpa.2022.102199>

Zhao et al., 2023 – Zhao P., Feng Y., Wu J., Zhu J., Yang J., Ma X., Ouyang Z., Zhang X., Zhang W., Wang W. Efficient sample preparation system for multi-omics analysis via single cell mass spectrometry // *Analytical Chemistry*. 2023. 95(18): 7212-7219. DOI: <https://doi.org/10.1021/acs.analchem.2c05728>

Ziech et al., 2011 – Ziech D., Franco R., Pappa A., Panayiotidis M.I. Reactive Oxygen Species (ROS) – Induced genetic and epigenetic alterations in human carcinogenesis // *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis*. 2011. 711(1-2): 167-173. DOI: <https://doi.org/10.1016/j.mrfmmm.2011.02.0150>

Ziech et al., 2012 – Ziech D., Anestopoulos I., Hanafi R., Voulgaridou G.P., Franco R., Georgakilas A.G., Pappa A., Panayiotidis M.I. Pleiotrophic effects of natural products in ROS-induced carcinogenesis: the role of plant-derived natural products in oral cancer chemoprevention // *Cancer Letters*. 2012. 327(1-2): 16-25. DOI: <https://doi.org/10.1016/j.canlet.2012.02.025>

References

Andrews et al., 1979 – Andrews, E.J., Todd, P.W., Kukulinsky, N.E. (1979). Surface charge in foreign body carcinogenesis. *Journal of Biomedical Materials Research*. 13(2): 173-187. DOI: <10.1002/jbm.820130203>

[Antonios et al., 2022](#) – Antonios M.A., Raouf M.M., Ghoniem S.A., Hassan, E.H. (2022). Clinical Impact of Biotyping of Klebsiella pneumoniae Isolates From Health Care–Associated Infections Using MALDI-TOF-MS. *Infectious Diseases in Clinical Practice*. 30(4): e1143. DOI: 10.1097/IPC.oooooooooooo0001143

[Arthur et al., 2017](#) – Arthur, K.L., Turner, M.A., Reynolds, J.C., Creaser, C.S. (2017). Increasing peak capacity in nontargeted omics applications by combining full scan field asymmetric waveform ion mobility spectrometry with liquid chromatography–mass spectrometry. *Analytical chemistry*. 89(6): 3452-3459. DOI: 10.1021/acs.analchem.6bo4315

[Asada et al., 2018](#) – Asada, M., Hakimi, H., Kawazu, S.I. (2018). The application of the HyPer fluorescent sensor in the real-time detection of H₂O₂ in Babesia bovis merozoites in vitro. *Veterinary parasitology*. 255: 78-82. DOI: <https://doi.org/10.1016/j.vetpar.2018.03.016>

[Bagatolli et al., 2021](#) – Bagatolli, L.A., Mangiarotti, A., Stock, R.P. (2021). Cellular metabolism and colloids: Realistically linking physiology and biological physical chemistry. *Progress in Biophysics and Molecular Biology*. 162: 79-88. DOI: <https://doi.org/10.1016/j.pbiomolbio.2020.06.002>

[Banerjee et al., 2023](#) – Banerjee, S., Hatimuria, M., Sarkar, K., Das, J., Pabbathi, A., Sil, P.C. (2023). Recent Contributions of Mass Spectrometry-Based “Omics” in the Studies of Breast Cancer. *Chemical Research in Toxicology*. 37(2): 137-180. DOI: <https://doi.org/10.1021/acs.chemrestox.3c00223>

[Beech, 1989a](#) – Beech, J.A. (1989). The membrane potential theory of carcinogenesis. *Medical Hypotheses*. 29(2): 101-104. DOI: [https://doi.org/10.1016/0306-9877\(89\)90069-8](https://doi.org/10.1016/0306-9877(89)90069-8)

[Beech, 1989b](#) – Beech, J.A. (1989). Two stage carcinogenesis by membrane potential changes. *Medical Hypotheses*. 29(3): 217-221. DOI: [https://doi.org/10.1016/0306-9877\(89\)90196-5](https://doi.org/10.1016/0306-9877(89)90196-5)

[Beech, 1994](#) – Beech, J.A. (1994). Carcinogenesis and initiation of cell cycling by charge-induced membrane clusters may be due to mitogen receptors and Na⁺ H⁺ antiports. *Medical hypotheses*. 42(6): 385-389. DOI: [https://doi.org/10.1016/0306-9877\(94\)90158-9](https://doi.org/10.1016/0306-9877(94)90158-9)

[Begum, Shen, 2023](#) – Begum, H.M., Shen, K. (2023). Intracellular and microenvironmental regulation of mitochondrial membrane potential in cancer cells. *WIREs mechanisms of disease*. 15(3): e1595. DOI: <https://doi.org/10.1002/wsbm.1595>

[Belizario et al., 2015](#) – Belizario, J.E., Sangiliano, B.A., Perez-Sosa, M., Santos, B.V., Machado-Santelli, G.M. (2015). Advances in the integration of optical and mass spectrometry molecular imaging technologies: from omics data to molecular signature discovery. *Discovery Medicine*. 20(112): 393-401.

[Belousov et al., 2006](#) – Belousov, V.V., Fradkov, A.F., Lukyanov, K.A., Staroverov, D.B., Shakhbazov, K.S., Terskikh, A.V., Lukyanov, S. (2006). Genetically encoded fluorescent indicator for intracellular hydrogen peroxide. *Nature methods*. 3(4): 281-286. DOI: <https://doi.org/10.1038/nmeth866>

[Berrazeg et al., 2013](#) – Berrazeg, M., Diene, S. M., Drissi, M., Kempf, M., Richet, H., Landraud, L., Rolain, J.M. (2013). Biotyping of multidrug-resistant Klebsiella pneumoniae clinical isolates from France and Algeria using MALDI-TOF MS. *PloS one*. 8(4): e61428. DOI: <https://doi.org/10.1371/journal.pone.0061428>

[Bilan, Belousov, 2016](#) – Bilan, D.S., Belousov, V.V. (2016). HyPer family probes: state of the art. *Antioxidants & redox signaling*. 24(13): 731-751. DOI: <https://doi.org/10.1089/ars.2015.6586>

[Bilbao et al., 2021](#) – Bilbao, A., Gibbons, B.C., Stow, S.M., Kyle, J.E., Bloodsworth, K.J., Payne, S.H., Smith, R.D., Ibrahim, Y.M., Baker, E.S., Fjeldsted, J.C. (2021). A preprocessing tool for enhanced ion mobility–mass spectrometry-based omics workflows. *Journal of proteome research*. 21(3): 798-807. DOI: <https://doi.org/10.1021/acs.jproteome.1c00425>

[Bocklitz et al., 2013](#) – Bocklitz, T.W., Crecelius, A.C., Matthaus, C., Tarcea, N., von Eggeling, F., Schmitt, M., Schubert, U.S., Popp, J. (2013). Deeper understanding of biological tissue: quantitative correlation of MALDI-TOF and Raman imaging. *Analytical chemistry*. 85(22): 10829-10834. DOI: <https://doi.org/10.1021/ac402175c>

[Bocklitz et al., 2015](#) – Bocklitz, T., Bräutigam, K., Urbanek, A., Hoffmann, F., von Eggeling, F., Ernst, G., Schmitt, M., Schubert, U., Guntinas-Lichius, O., Popp, J. (2015). Novel workflow for combining Raman spectroscopy and MALDI-MSI for tissue based studies. *Analytical and bioanalytical chemistry*. 407: 7865-7873. DOI: <https://doi.org/10.1007/s00216-015-8987-5>

- Bonham et al., 2014** – Bonham, C.A., Steevensz, A.J., Geng, Q., Vacratsis, P.O. (2014). Investigating redox regulation of protein tyrosine phosphatases using low pH thiol labeling and enrichment strategies coupled to MALDI-TOF mass spectrometry. *Methods*. 65(2): 190-200. DOI: <https://doi.org/10.1016/j.ymeth.2013.08.014>
- Boyer et al., 2017** – Boyer, P.H., Boulanger, N., Nebbak, A., Collin, E., Jaulhac, B., Almeras, L. (2017). Assessment of MALDI-TOF MS biotyping for Borrelia burgdorferi sl detection in Ixodes ricinus. *PLoS One*. 12(9): e0185430. DOI: <https://doi.org/10.1371/journal.pone.0185430>
- Brilkina et al., 2018** – Brilkina, A.A., Peskova, N.N., Dudenkova, V.V., Gorokhova, A.A., Sokolova, E.A., Balalaeva, I.V. (2018). Monitoring of hydrogen peroxide production under photodynamic treatment using protein sensor HyPer. *Journal of Photochemistry and Photobiology B: Biology*. 178: 296-301. DOI: <https://doi.org/10.1016/j.jphotobiol.2017.11.020>
- Burioni et al., 2024** – Burioni, R., Silvestrini, L., D'Orto, B., Tetè, G., Nagni, M., Polizzi, E., Gherlone, E.F. (2024). Could Dental Material Reuse Play a Significant Role in Preservation of Raw Materials, Water, Energy, and Costs? Microbiological Analysis of New versus Reused Healing Abutments: An In Vitro Study. *Bioengineering*. 11(4): 387. DOI: <https://doi.org/10.3390/bioengineering11040387>
- Butterfield et al., 2014** – Butterfield, D.A., Gu, L., Domenico, F.D., Robinson, R.A. (2014). Mass spectrometry and redox proteomics: applications in disease. *Mass spectrometry reviews*. 33(4): 277-301. DOI: <https://doi.org/10.1002/mas.21374>
- Bykova, Rampitsch, 2013** – Bykova, N.V., Rampitsch, C. (2013). Modulating protein function through reversible oxidation: redox-mediated processes in plants revealed through proteomics. *Proteomics*. 13(3-4): 579-596. DOI: <https://doi.org/10.1002/pmic.201200270>
- Bykova et al., 2011** – Bykova, N.V., Hoehn, B., Rampitsch, C., Banks, T., Stebbing, J.A., Fan, T., Knox, R. (2011). Redox-sensitive proteome and antioxidant strategies in wheat seed dormancy control. *Proteomics*. 11(5): 865-882. DOI: <https://doi.org/10.1002/pmic.200900810>
- Byrne, 2024** – Byrne, H.J. (2024). Spectralomics—Towards a holistic adaptation of label free spectroscopy. *Vibrational Spectroscopy*. 132: 103671. DOI: <https://doi.org/10.1016/j.vibspec.2024.103671>
- Causon et al., 2020** – Causon, T.J., Kurulugama, R.T., Hann, S. (2020). Drift-tube ion mobility-mass spectrometry for nontargeted' omics. *Methods in Molecular Biology*. 2084: 79-94. DOI: https://doi.org/10.1007/978-1-0716-0030-6_4
- Cavalieri, Calvin, 1972** – Cavalieri, E., Calvin, M. (1972). Charge localization in carbonium-ion of methylbenzanthracenes-clue to their mechanism of carcinogenesis. *Proceedings of the American Association for Cancer Research*. 13(MAR): 125.
- Challen, Cramer, 2022** – Challen, B., Cramer, R. (2022). Advances in ionisation techniques for mass spectrometry-based omics research. *Proteomics*. 22(15-16): 2100394. DOI: <https://doi.org/10.1002/pmic.202100394>
- Chao, Zongwei, 2021** – Chao, Z., Zongwei, C. (2021). Mass spectrometry imaging and omics for environmental toxicology research. *Progress in chemistry*. 33(4): 503. DOI: <https://doi.org/10.1016/j.cbpa.2022.102199>
- Chouchane, 1996** – Chouchane, S. (1996). Etude des proprietes electrophiles generees en milieu aqueux aere, par des poussières d'hematite ou de magnetite. Relation probable avec le cancer bronchopulmonaire (Doctoral dissertation, Paris 6). [Electronic resource]. URL: <https://theses.fr/1992PA066268>
- Chudakov et al., 2010** – Chudakov, D.M., Matz, M.V., Lukyanov, S., Lukyanov, K.A. (2010). Fluorescent proteins and their applications in imaging living cells and tissues. *Physiological reviews*. 90(3): 1103-1163. DOI: <https://doi.org/10.1152/physrev.00038.2009>
- Cornellison et al., 2011** – Cornellison, C.D., Dyer, J.M., Plowman, J.E., Krsinic, G.L., Clerens, S. (2011). MALDI-MS redox lipidomics applied to human hair: A first look. *International Journal of Trichology*. 3(1): 25-27. DOI: <https://doi.org/10.4103/0974-7753.82127>
- Danes et al., 2021** – Danes, J.M., Palma, F.R., Bonini, M.G. (2021). Arsenic and other metals as phenotype driving electrophiles in carcinogenesis. *Seminars in Cancer Biology*. 76: 287-291. DOI: <https://doi.org/10.1016/j.semcan.2021.09.012>
- Delafield et al., 2022** – Delafield, D.G., Lu, G., Kaminsky, C.J., Li, L. (2022). High-end ion mobility mass spectrometry: A current review of analytical capacity in omics applications and structural

investigations. *TrAC Trends in Analytical Chemistry*. 157: 116761. DOI: <https://doi.org/10.1016/j.trac.2022.116761>

DeLaney et al., 2018 – DeLaney, K., Sauer, C.S., Vu, N.Q., Li, L. (2018). Recent advances and new perspectives in capillary electrophoresis-mass spectrometry for single cell “omics”. *Molecules*. 24(1): 42. DOI: <https://doi.org/10.3390/molecules24010042>

Delfosse et al., 2016 – Delfosse, K., Wozny, M.R., Jaipargas, E.A., Barton, K.A., Anderson, C., Mathur, J. (2016). Fluorescent protein aided insights on plastids and their extensions: a critical appraisal. *Frontiers in Plant Science*. 6: 1253. DOI: <https://doi.org/10.3389/fpls.2015.01253>

Delisi et al., 2024 – Delisi, D., Eskandari, N., Gentile, S. (2024). Membrane potential: A new hallmark of cancer. *Advances in cancer research*. 164: 93-110. DOI: <https://doi.org/10.1016/bs.acr.2024.04.010>

DePaoli et al., 2020 – DePaoli, D., Lemoine, É., Ember, K., Parent, M., Prud'homme, M., Cantin, L., Petrecca, K., Leblond, F., Côté, D.C. (2020). Rise of Raman spectroscopy in neurosurgery: a review. *Journal of biomedical optics*. 25(5): 050901. DOI: <https://doi.org/10.1117/1.jbo.25.5.050901>

Deweza et al., 2019 – Dewez, F., Martin-Lorenzo, M., Herfs, M., Baiwir, D., Mazzucchelli, G., De Pauw, E., Heeren, R.M., Balluff, B. (2019). Precise co-registration of mass spectrometry imaging, histology, and laser microdissection-based omics. *Analytical and bioanalytical chemistry*. 411: 5647-5653. DOI: <https://doi.org/10.1007/s00216-019-01983-z>

Donohoe, 2016 – Donohoe, G.C. (2016). Expanding the Applications of Ion Mobility Spectrometry and Mass Spectrometry in Integrative'Omics Analyses. PhD Dissertation, West Virginia University. DOI: <https://doi.org/10.33915/etd.5505>

Dunnington et al., 2024 – Dunnington, E.L., Wong, B.S., Fu, D. (2024). Innovative Approaches for Drug Discovery: Quantifying Drug Distribution and Response with Raman Imaging. *Analytical Chemistry*. 96(20): 7926–7944. DOI: <https://doi.org/10.1021/acs.analchem.4c01413>

Emerit, 1994 – Emerit, I. (1994). Reactive oxygen species, chromosome mutation, and cancer: possible role of clastogenic factors in carcinogenesis. *Free Radical Biology and Medicine*. 16(1): 99-109. DOI: [https://doi.org/10.1016/0891-5849\(94\)90246-1](https://doi.org/10.1016/0891-5849(94)90246-1)

Fernandez-Garcia, Olmos, 2014 – Fernandez-Garcia, N., Olmos, E. (2014). ROS and NOS imaging using microscopical techniques. *Plant Image Analysis: Fundamentals and Applications*. 245. DOI: <http://dx.doi.org/10.1201/b17441-13>

Fischer et al. 1987 – Fischer, S.M., Cameron, G.S., Baldwin, J.K., Jascheway, D.W., Patrick, K.E. (1988). Reactive oxygen in the tumor promotion stage of skin carcinogenesis. *Lipids*. 23(6): 592-597. DOI: <https://doi.org/10.1007/bf02535603>

Fischer, 1987 – Fischer, S. (1987). Reactive oxygen in the tumor promotion stage of carcinogenesis. *Journal of the American Oil Chemists Society*. 64(5): 630.

Forrest, 2015 – Forrest, M.D. (2015). Why cancer cells have a more hyperpolarised mitochondrial membrane potential and emergent prospects for therapy. *BioRxiv*. 025197. DOI: <https://doi.org/10.1101/025197>

Franceschi et al., 2013 – Franceschi, P., Giordan, M., Wehrens, R. (2013). Multiple comparisons in mass-spectrometry-based-omics technologies. *TrAC Trends in Analytical Chemistry*. 50: 11-21. DOI: <https://doi.org/10.1016/j.trac.2013.04.011>

Frey-Wyssling, 1965 – Frey-Wyssling, A. (1965). Comparative organellography. *Experientia*. 21(12): 681-687. DOI: <https://doi.org/10.1007/bf02138470>

Friday et al., 2011 – Friday, E., Oliver III, R., Welbourne, T., Turturro, F. (2011). Glutaminolysis and glycolysis regulation by troglitazone in breast cancer cells: relationship to mitochondrial membrane potential. *Journal of cellular physiology*. 226(2): 511-519. DOI: <https://doi.org/10.1002/jcp.22360>

Gąbka et al., 2021 – Gąbka, M., Dalek, P., Przybyło, M., Gackowski, D., Oliński, R., Langner, M. (2021). The membrane electrical potential and intracellular pH as factors influencing intracellular ascorbate concentration and their role in cancer treatment. *Cells*. 10(11): 2964. DOI: <https://doi.org/10.3390/cells10112964>

Gayan et al., 2022 – Gayan, S., Joshi, G., Dey, T. (2022). Biomarkers of mitochondrial origin: a futuristic cancer diagnostic. *Integrative Biology*. 14(4): 77-88. DOI: <https://doi.org/10.1093/intbio/zyac008>

Gekenidis et al., 2014 – Gekenidis, M.T., Studer, P., Wüthrich, S., Brunisholz, R., Drissner, D. (2014). Beyond the matrix-assisted laser desorption ionization (MALDI) biotyping workflow: in search

of microorganism-specific tryptic peptides enabling discrimination of subspecies. *Applied and environmental microbiology*. 80(14): 4234-4241. DOI: <https://doi.org/10.1128/aem.00740-14>

Girolamo et al., 2013 – Girolamo, F.D., Lante, I., Muraca, M., Putignani, L. (2013). The role of mass spectrometry in the “omics” era. *Current organic chemistry*. 17(23): 2891-2905. DOI: <https://doi.org/10.2174/1385272817888131118162725>

Gobert et al., 2021 – Gobert, A.P., Boutaud, O., Asim, M., Zagol-Ikapitte, I.A., Delgado, A.G., Latour, Y.L., Finley, J.L., Singh, K., Verriere, T.G., Allaman, M.M., Barry, D.P. (2021). Dicarbonyl electrophiles mediate inflammation-induced gastrointestinal carcinogenesis. *Gastroenterology*. 160(4): 1256-1268. DOI: <https://doi.org/10.1053/j.gastro.2020.11.006>

Gogichadze et al., 2014 – Gogichadze, G., Gogichadze, T., Misabishvili, E., Kamkamidze, G. (2014). Possible effect of variable membrane potential of a cancer cell on different carcinogenic processes. *Georgian medical news*. (234): 116-120.

Gokulan et al., 2020 – Gokulan, R.C., Paulrasu, K., Zaika, E., Palrasu, M., Boutaud, O., Dikalov, S., Zaika, A. (2020). SU1171 - NADPH oxidases and reactive oxygen species generates isolog protein adducts in the esophagus: a novel mechanism that triggers esophageal carcinogenesis. *Gastroenterology*. 158(6): S-531.

Gradov, 2012 – Gradov, O.V. (2012). Eksperimental'nye ustavovki dlya ozonometricheskoi mikroskopii [Experimental setups for ozonometric microscopy]. *Meditinskaya tekhnika*. (6): 42-47. [in Russian]

Gradov, 2013 – Gradov, O.V. (2013). Experimental Setups for Ozonometric Microscopy. *Biomedical Engineering*. 46(6): 260-264. DOI: <http://dx.doi.org/10.1007/s10527-013-9319-8>

Grigorov, 2012 – Grigorov, B. (2012). Reactive oxygen species and their relation to carcinogenesis. *Trakia journal of sciences*. 10(3): 83-92.

Guerreiro et al., 2024 – Guerreiro, E.M., Kruglik, S.G., Swamy, S., Latysheva, N., Østerud, B., Guigner, J.M., Sureau, F., Bonneau, S., Kuzmin, A.N., Prasad, P.N., Hansen, J.B. (2024). Extracellular vesicles from activated platelets possess a phospholipid-rich biomolecular profile and enhance prothrombinase activity. *Journal of Thrombosis and Haemostasis*. 22(5): 1463-1474. DOI: <https://doi.org/10.1016/j.jtha.2024.01.004>

Hadacek, Bachmann, 2015 – Hadacek, F., Bachmann, G. (2015). Low-molecular-weight metabolite systems chemistry. *Frontiers in environmental science*. 3: 12. DOI: <https://doi.org/10.3389/fenvs.2015.00012>

Harach et al., 2019 – Harach, J.L., Lynch, C.J., Montovano, G., Olsen, S.H., Foley, T.D. (2019). Glyceraldehyde-3-phosphate dehydrogenase differentiates the cytotoxic mechanisms of cancer-active electrophiles in a yeast model. *The FASEB Journal*. 33(S1): 652-628. DOI: http://dx.doi.org/10.1096/fasebj.2019.33.1_supplement.652.8

Hong et al., 2021 – Hong, J.K., Kim, S.B., Lyou, E.S., Lee, T.K. (2021). Microbial phenomics linking the phenotype to function: The potential of Raman spectroscopy. *Journal of Microbiology*. 59: 249-258. DOI: <https://doi.org/10.1007/s12275-021-0590-1>

Houdelet, 2015 – Houdelet, C., Bocquet, M., Bulet, P. (2020). MALDI Biotyping, an approach for deciphering and assessing the identity of the honeybee pathogen Nosema. *Rapid Communications in Mass Spectrometry*. 35(3): e8980. DOI: <http://dx.doi.org/10.1002/rcm.8980>

Huang, Spiers, 2006 – Huang, W.E., Spiers, A.J. (2006). Consideration of future requirements for Raman microbiology as an exemplar for the ab initio development of informatics frameworks for emergent OMICS technologies. *OMICS: A Journal of Integrative Biology*. 10(2): 238-241. DOI: <https://doi.org/10.1089/omi.2006.10.238>

Huang et al., 1994 – Huang, X., Zhuang, Z., Frenkel, K., Klein, C. B., Costa, M. (1994). The role of nickel and nickel-mediated reactive oxygen species in the mechanism of nickel carcinogenesis. *Environmental health perspectives*. 102(Suppl 3): 281-284. DOI: <https://doi.org/10.1289/ehp.94102s3281>

Huang et al., 2023 – Huang, L., Sun, H., Sun, L., Shi, K., Chen, Y., Ren, X., Ge, Y., Jiang, D., Liu, X., Knoll, W., Zhang, Q. (2023). Rapid, label-free histopathological diagnosis of liver cancer based on Raman spectroscopy and deep learning. *Nature Communications*. 14(1): 48. DOI: <https://doi.org/10.1038/s41467-022-35696-2>

Jablokov, Gradov, 2016 – Gradov, O.V., Jablokov, A.G. (2016). Novel morphometrics-on-a-chip: CCD- or CMOS-lab-on-a-chip based on discrete converters of different physical and chemical parameters of histological samples into the optical signals with positional sensitivity for

morphometry of non-optical patterns. *Journal of Biomedical Technologies*. (2): 1-29. DOI: <http://dx.doi.org/10.15393/j6.art.2016.3642>

Jablokow, Gradow, 2015a – *Jablokow, A.G., Gradow, O.V.* (2015). MS-FRAP or MALDI Imaging Setups With Programmable Laser Sources: a New Way to the Diffusion, Molecular Mobility and Binding Measurements. In: Progr. 63-rd ASMS Conference on Mass Spectrometry and Allied Topics (St. Louis; June 2015), Section: "Imaging MS: Instrumentation" (JASMS Special Issue). DOI: [10.13140/RG.2.1.4919.1841](https://doi.org/10.13140/RG.2.1.4919.1841)

Jablokow, Gradow, 2015b – *Jablokow, A., Gradow, O.* (2015). Verifying Continuity of Membranous Organelles and Measurements of Exchange Rate Between the Nucleus and Cytoplasm using FLIP-Like MALDI-Based Imaging. In: Progr. 63-rd ASMS Conference on Mass Spectrometry and Allied Topics (St. Louis; June 2015), Section: "Imaging MS: Instrumentation" (JASMS Special Issue). DOI: [10.13140/RG.2.1.2322.3203](https://doi.org/10.13140/RG.2.1.2322.3203)

Jablokow et al., 2017 – *Jablokow, A.G., Skrynnik, A.A., Orekhov, F.K., Nasirov, P.A., Grakov, O.V.* (2017). "MALDI-FLIP-on-a-chip" and " MALDI-FRAP-on-a-flap": Novel Techniques for Soil Microbiology and Environmental Biogeochemistry. I-MALDI Chip Fingerprinting. *Biogeosystem Technique*. (4): 140-188. DOI: <https://doi.org/10.13187/bgt.2017.2.140>

Jablokow et al., 2018 – *Jablokow, A.G., Nasirov, P.A., Orekhov, F.K., Grakov, O.V.* (2018). "MALDI-FLIP-on-a-chip" and " MALDI-FRAP-on-a-flap": Novel Techniques for Soil Microbiology and Environmental Biogeochemistry. II-Polymer Chip Prototyping. *Biogeosystem Technique*. (5): 3-56. DOI: <https://doi.org/10.13187/bgt.2018.1.3>

Jaeken, 2017 – *Jaeken, L.* (2017). The neglected functions of intrinsically disordered proteins and the origin of life. *Progress in Biophysics and Molecular Biology*. 126: 31-46. DOI: <https://doi.org/10.1016/j.pbiomolbio.2017.03.002>

Janků et al., 2019a – *Janků, M., Luhová, L., Petřivalský, M.* (2019). On the origin and fate of reactive oxygen species in plant cell compartments. *Antioxidants*. 8(4): 105. DOI: <https://doi.org/10.3390/antiox8040105>

Janků et al., 2019b – *Janků, M., Tichá, T., Luhová, L., Petřivalský, M.* (2019). Chapter 40: Compartmentalization of reactive Oxygen species and Nitric Oxide production in plant cells. *Reactive oxygen, nitrogen and sulfur species in plants: production, Metabolism, Signaling and Defense Mechanisms*, 923-945. DOI: <https://doi.org/10.1002/9781119468677.ch40>

Jian, 2023 – *Jian, X.U.* (2023). Ramanome, FlowRACS and RACS-Seq/Culture: functional dissection and mining of microbiomes at single-cell resolution. [Electronic resource]. URL: https://www.ramanfestconf.com/2023/Abstracts/2023_Xu.pdf

Jones, Sies, 2015 – *Jones, D.P., Sies, H.* (2015). The redox code. *Antioxidants & redox signaling*. 23(9): 734-746. DOI: <https://doi.org/10.1089/ars.2015.6247>

Jopkiewicz, 2019 – *Jopkiewicz, S.P.* (2019). Significance of reactive oxygen species and oxidative stress in carcinogenesis. *Journal of Education, Health and Sport*. 9(6): 241-250. DOI: <http://dx.doi.org/10.5281/zenodo.3243595>

Klein, Costa, 1991 – *Klein, C.B., Costa, M.* (1991). The Role of Nickel and Nickel-mediated Reactive Oxygen Species in the Mechanism of Nickel Carcinogenesis. *Environmental Health Perspectives*. 102: 281. DOI: <https://doi.org/10.1289/ehp.94102s3281>

Kosmachevskaya, Topunov, 2021 – *Kosmachevskaya, O.V., Topunov, A.F.* (2021). Nonenzymatic reactions in metabolism: Their role in evolution and adaptation. *Applied Biochemistry and Microbiology*. 57: 543-555. DOI: <http://dx.doi.org/10.1134/S0003683821050100>

Kovacic, Abadjian, 2018 – *Kovacic, P., Abadjian, M.C.Z.* (2018). Mechanism of lung carcinogenesis: electron transfer, reactive oxygen species, oxidative stress and antioxidants. *SOJ Microbiol Infect Dis*. 2: 1-10.

Kovacic et al., 1986 – *Kovacic, P., Crawford, P.W., Ryan, M.D., Nelson, V.C.* (1986). 844– Charge transfer mechanism for carcinogenesis by alkylating and other agents. *Bioelectrochemistry and Bioenergetics*. 15(3): 305-316. DOI: [https://doi.org/10.1016/0302-4598\(86\)85020-6](https://doi.org/10.1016/0302-4598(86)85020-6)

Kruk, Aboul-Enein, 2017 – *Kruk, J., Aboul-Enein, H.* (2017). Reactive oxygen and nitrogen species in carcinogenesis: implications of oxidative stress on the progression and development of several cancer types. *Mini reviews in medicinal chemistry*. 17(11): 904-919. DOI: <https://doi.org/10.2174/1389557517666170228115324>

Kuwahara et al., 2021 – *Kuwahara, Y., Tomita, K., Roudkenar, M.H., Roushandeh, A.M., Urushihara, Y., Igarashi, K., Kurimasa, A., Sato, T.* (2021). Decreased mitochondrial membrane

potential is an indicator of radioresistant cancer cells. *Life Sciences*. 286: 120051. DOI: <https://doi.org/10.1016/j.lfs.2021.120051>

Kuzmin et al., 2017a – Kuzmin, A. N., Pliss, A., Prasad, P.N. (2017). Ramanomics: new omics disciplines using micro Raman spectrometry with biomolecular component analysis for molecular profiling of biological structures. *Biosensors*. 7(4): 52. DOI: <https://doi.org/10.3390/bios7040052>

Kuzmin et al., 2017b – Kuzmin, A.N., Levchenko, S.M., Pliss, A., Qu, J., Prasad, P.N. (2017). Molecular profiling of single organelles for quantitative analysis of cellular heterogeneity. *Scientific Reports*. 7(1): 6512. DOI: <https://doi.org/10.1038/s41598-017-06936-z>

Kuzmin et al., 2018 – Kuzmin, A.N., Pliss, A., Rzhevskii, A., Lita, A., Larion, M. (2018). BCABox algorithm expands capabilities of Raman microscope for single organelles assessment. *Biosensors*. 8(4): 106. DOI: <https://doi.org/10.3390/bios8040106>

LaLone et al., 2023 – LaLone, V., Aizenshtadt, A., Goertz, J., Skottvoll, F.S., Mota, M.B., You, J., Zhao, X., Berg, H.E., Stokowiec, J., Yu, M., Schwendeman, A. (2023). Quantitative chemometric phenotyping of three-dimensional liver organoids by Raman spectral imaging. *Cell Reports Methods*. 3(4): 100440. DOI: <https://doi.org/10.1016/j.crmeth.2023.100440>

LaLone et al., 2019a – LaLone, V., Fawaz, M.V., Morales-Mercado, J., Mourão, M.A., Snyder, C.S., Kim, S.Y., Lieberman, A.P., Tuteja, A., Mehta, G., Standiford, T.J., Raghavendran, K. (2019). Inkjet-printed micro-calibration standards for ultraquantitative Raman spectral cytometry. *Analyst*. 144(12): 3790-3799. DOI: <https://doi.org/10.1039/c9an00500e>

LaLone et al., 2019b – LaLone, V., Mourão, M.A., Standiford, T.J., Raghavendran, K., Shedd, K., Stringer, K.A., Rosania, G.R. (2019). An expandable mechanopharmaceutical device (3): A versatile Raman spectral cytometry approach to study the drug cargo capacity of individual macrophages. *Pharmaceutical Research*. 36(1):2. DOI: <https://doi.org/10.1007/s11095-018-2540-0>

Lareau, 2016 – Lareau, N.M. (2016). Development of Ion Mobility and Mass Spectrometry Strategies in Support of Integrated Omics and Systems Biology. Vanderbilt University.

Larion et al., 2018 – Larion, M., Dowdy, T., Ruiz-Rodado, V., Meyer, M.W., Song, H., Zhang, W., Davis, D., Gilbert, M.R., Lita, A. (2018). Detection of metabolic changes induced via drug treatments in live cancer cells and tissue using Raman imaging microscopy. *Biosensors*. 9(1): 5. DOI: <https://doi.org/10.3390/bios9010005>

Lawrence, 2023 – Lawrence, C.P. (2023). Simple machine learning methods work surprisingly well for Ramanomics. *Journal of Raman Spectroscopy*. 54(8): 887-889. DOI: <https://doi.org/10.1002/jrs.6555>

Lee et al., 2016 – Lee, D.G., Choi, B.K., Kim, Y.H., Oh, H.S., Park, S.H., Bae, Y.S., Kwon, B.S. (2016). The repopulating cancer cells in melanoma are characterized by increased mitochondrial membrane potential. *Cancer letters*. 382(2): 186-194. DOI: <https://doi.org/10.1016/j.canlet.2016.08.027>

Lei et al., 2021 – Lei, L., Zhang, J., Decker, E.A., Zhang, G. (2021). Roles of lipid peroxidation-derived electrophiles in pathogenesis of colonic inflammation and colon cancer. *Frontiers in Cell and Developmental Biology*. 9: 665591. DOI: <https://doi.org/10.3389/fcell.2021.665591>

Lemeshko, 2015 – Lemeshko, V.V. (2015). Channeling of mitochondrial energy in cardiac and cancer cells by the metabolically-dependent outer membrane potential. *Biophysical Journal*. 108(2): 607a.

Lennicke et al., 2016 – Lennicke, C., Rahn, J., Heimer, N., Lichtenfels, R., Wessjohann, L.A., Seliger, B. (2016). Redox proteomics: Methods for the identification and enrichment of redox-modified proteins and their applications. *Proteomics*. 16(2): 197-213. DOI: <https://doi.org/10.1002/pmic.201500268>

Li et al., 2020 – Li, X., Yang, F., Rubinsky, B. (2020). A correlation between electric fields that target the cell membrane potential and dividing HeLa cancer cell growth inhibition. *IEEE Transactions on Biomedical Engineering*. 68(6): 1951-1956. DOI: <https://doi.org/10.1109/tbme.2020.3042650>

Lorenz et al., 2020 – Lorenz, B., Ali, N., Bocklitz, T., Rösch, P., Popp, J. (2020). Discrimination between pathogenic and non-pathogenic *E. coli* strains by means of Raman microspectroscopy. *Analytical and Bioanalytical Chemistry*. 412: 8241-8247. DOI: <https://doi.org/10.1007/s00216-020-02957-2>

Lu et al., 2020 – Lu, W., Chen, X., Wang, L., Li, H., Fu, Y.V. (2020). Combination of an artificial intelligence approach and laser tweezers Raman spectroscopy for microbial identification. *Analytical chemistry*. 92(9): 6288-6296. DOI: <https://doi.org/10.1021/acs.analchem.9b04946>

Lu et al., 2023 – Lu, H., Zhang, H., Li, L. (2023). Chemical tagging mass spectrometry: an approach for single-cell omics. *Analytical and bioanalytical chemistry*. 415(28): 6901-6913. DOI: <https://doi.org/10.1007/s00216-023-04850-0>

Luo et al., 2022 – Luo, Y., Sobhani, Z., Zhang, Z., Zhang, X., Gibson, C.T., Naidu, R., Fang, C. (2022). Raman imaging and MALDI-MS towards identification of microplastics generated when using stationery markers. *Journal of Hazardous Materials*. 424: 127478. DOI: <https://doi.org/10.1016/j.jhazmat.2021.127478>

Lyublinskaya, Antunes, 2019 – Lyublinskaya, O., Antunes, F. (2019). Measuring intracellular concentration of hydrogen peroxide with the use of genetically encoded H₂O₂ biosensor HyPer. *Redox biology*. 24: 101200. DOI: <https://doi.org/10.1016/j.redox.2019.101200>

MacLeod et al., 2009 – MacLeod, A.K., McMahon M., Plummer, S.M., Higgins, L.G., Penning, T.M., Igarashi, K., Hayes, J.D. (2009). Characterization of the cancer chemopreventive NRF2-dependent gene battery in human keratinocytes: demonstration that the KEAP1–NRF2 pathway, and not the BACH1–NRF2 pathway, controls cytoprotection against electrophiles as well as redox-cycling compounds. *Carcinogenesis*. 30(9): 1571-1580. DOI: <https://doi.org/10.1093/carcin/bgp176>

Mallah et al., 2023 – Mallah, K., Zibara, K., Kerbaj, C., Eid, A., Khoshman, N., Ousseily, Z., Kobeissy, A., Cardon, T., Cizkova, D., Kobeissy, F., Fournier, I. (2023). Neurotrauma investigation through spatial omics guided by mass spectrometry imaging: Target identification and clinical applications. *Mass spectrometry reviews*. 42(1): 189-205. DOI: <https://doi.org/10.1002/mas.21719>

Marino et al., 1994 – Marino, A.A., Iliev, I.G., Schwalke, M.A., Gonzalez, E., Marler, K.C., Flanagan, C.A. (1994). Association between cell membrane potential and breast cancer. *Tumor Biology*. 15(2): 82-89. DOI: <https://doi.org/10.1159/000217878>

Marquez-Quinones, 2007 – Marquez-Quinones, A. (2007). *Reactive oxygen species, hepatitis and carcinogenesis initiation: an integrative approach combining transcriptomic and metabonomic profilings* (Doctoral dissertation, Institut National des Sciences Appliquées de Toulouse). [Electronic resource]. URL: <https://theses.fr/2007ISAT0028>

Medeiros, 2018 – Medeiros, F.H.C. (2018). The role of reactive oxygen species in thyroid radio-carcinogenesis (Doctoral dissertation, Université Paris-Saclay (ComUE); Universidade federal do Rio de Janeiro). <https://theses.fr/2018SACLS085>. [Electronic resource]. URL: <https://theses.hal.science/tel-04010752> NNT : 2018SACLS085

Miller, 1998 – Miller, J.A. (1998). The metabolism of xenobiotics to reactive electrophiles in chemical carcinogenesis and mutagenesis: a collaboration with Elizabeth Cavert Miller and our associates. *Drug metabolism reviews*. 30(4): 645-674. DOI: <https://doi.org/10.3109/03602539808996326>

Moldogazieva et al., 2018 – Moldogazieva, N.T., Lutsenko, S.V., Terentiev, A.A. (2018). Reactive oxygen and nitrogen species-induced protein modifications: implication in carcinogenesis and anticancer therapy. *Cancer Research*. 78(21): 6040-6047. DOI: <https://doi.org/10.1158/0008-5472.can-18-0980>

Mosier-Boss, 2017 – Mosier-Boss, P.A. (2017). Review on SERS of Bacteria. *Biosensors*. 7(4): 51. DOI: <https://doi.org/10.3390/bios7040051>

Murray, Van Eyk, 2012 – Murray, C.I., Van Eyk, J.E. (2012). Chasing cysteine oxidative modifications: proteomic tools for characterizing cysteine redox status. *Circulation: Cardiovascular Genetics*. 5(5): 591. DOI: <https://doi.org/10.1161/CIRCGENETICS.111.961425>

Nie et al., 2016 – Nie, W., Yan, L., Lee, Y.H., Guha, C., Kurland, I.J., Lu, H. (2016). Advanced mass spectrometry-based multi-omics technologies for exploring the pathogenesis of hepatocellular carcinoma. *Mass Spectrometry Reviews*. 35(3): 331-349. DOI: <https://doi.org/10.1002/mas.21439>

Nishigori et al., 2004 – Nishigori, C., Hattori, Y., Toyokuni, S. (2004). Role of reactive oxygen species in skin carcinogenesis. *Antioxidants and Redox Signaling*. 6(3): 561-570. DOI: <https://doi.org/10.1089/152308604773934314>

Nnodim, Hauwa, 2020 – Nnodim, J., Hauwa, B. (2020). Membrane potential: An emerging and important player in cancer metastasis. *Asclep Med Res Rev*. 3: 1-2.

Okada, 2007 – Okada, F. (2007). Beyond foreign-body-induced carcinogenesis: impact of reactive oxygen species derived from inflammatory cells in tumorigenic conversion and tumor progression. *International journal of cancer*. 121(11): 2364-2372. DOI: <https://doi.org/10.1002/ijc.23125>

- Okazaki, 2022** – Okazaki, Y. (2022). Asbestos-induced mesothelial injury and carcinogenesis: Involvement of iron and reactive oxygen species. *Pathology International*. 72(2): 83-95. DOI: <https://doi.org/10.1111/pin.13196>
- Oliński, Jurgowiak, 1999** – Oliński, R., Jurgowiak, M. (1999). The role of reactive oxygen species in mutagenesis and carcinogenesis processes. *Postepy Biochemii*. 45(1): 50-58.
- Olsen et al., 2018** – Olsen, S., Foley, T., Montovano, G., Lynch, C. (2018). Use of phenylarsine oxide-affinity chromatography to identify common cellular targets of cancer-active electrophiles. *Free Radical Biology and Medicine*. 128: S72. DOI: <https://doi.org/10.1016/j.freeradbiomed.2018.10.154>
- Orekhov, Gradov, 2022** – Orekhov F., Gradov O. (2022). Automated Soil Microbiology Using Lensless and LDI MS Imaging with Buried Slides. *Smart Innovation, Systems and Technologies*. 247: 471-479. DOI: http://dx.doi.org/10.1007/978-981-16-3844-2_43
- Orekhov, Gradov, 2023** – Orekhov, F.K. Gradov, O.V. (2023). Towards Ultraviolet Microbeam Scanning and Lens-Less UV Microbeam Microscopy with Mirror Galvanometric Scanners: From the History of Research Instrumentation to Engineering of Modern Mechatronic Optical Systems. *J Sen Net Data Comm*. 3(1): 117-137.
- Orekhov, Gradov, 2023a** – Orekhov, F.K., Gradov, O.V. (2023). Target Chip Based Single-Cell Biotyping and Telemetric Bioluminescence Lensless Microscopy of the Buried Sandwich-Slides as a Novel Way for Measurement, Mapping and Molecular Imaging of Biodegradation / Biofouling of Plastic Surfaces in Real Soils. *Advances in Transdisciplinary Engineering*. 38: 417-425. DOI: <http://dx.doi.org/10.3233/ATDE230317>
- Orekhov, Gradov, 2023b** – Orekhov, T.K., Gradov, O.V. (2023). From desolvation-induced self-organization on the MALDI anchor target chip surfaces to laser-induced self-organization in MALDI techniques: Correlation-spectral analysis and complex wavelet analysis of tesiographic spots on the anchor chips. *Materials Technology Reports*. 1(1): 124. DOI: <http://dx.doi.org/10.59400/mtr.vi1.124>
- Orekhov et al., 2016** – Orekhov, F.K., Jablokow, A.G., Skrynnik, A.A. (2016). Hybridization of laser-induced spectrofluorescence analysis (LIFS), matrix-assisted laser desorption/ionization mass spectrometry (MALDI), fluorescence recovery after photobleaching (FRAP) and fluorescence loss in photobleaching (FLIP) microtechnics. *Journal of Biomedical Technologies*. (2): 42-52. DOI: <http://dx.doi.org/10.15393/j6.art.2016.3702>
- Orekhov et al., 2023** – Orekhov, F.K., Jablokow, A.G., Gradow, O.V. (2015). "Novel MALDI MS + FLIP approaches for verifying continuity of membranous structures and measurements of nucleus-cytoplasm exchange" (Poster). At: IMSIS 2023 (October 23-25, Montreal). Program and Abstracts. DOI: [10.13140/RG.2.2.31971.53287](https://doi.org/10.13140/RG.2.2.31971.53287)
- Pade et al., 2021** – Pade, L.R., Stepler, K.E., Portero, E.P., DeLaney, K., Nemes, P. (2024). Biological mass spectrometry enables spatiotemporal 'omics: From tissues to cells to organelles. *Mass spectrometry reviews*. 43(1): 106-138. DOI: <https://doi.org/10.1002/mas.21824>
- Paglia et al., 2022** – Paglia, G., Smith, A.J., Astarita, G. (2022). Ion mobility mass spectrometry in the omics era: Challenges and opportunities for metabolomics and lipidomics. *Mass Spectrometry Reviews*. 41(5): 722-765. DOI: <https://doi.org/10.1002/mas.21686>
- Panayiotidis, 2008** – Panayiotidis, M.I. (2008). Reactive oxygen species (ROS) in multistage carcinogenesis. *Cancer letters*. 266(1): 3-5. DOI: <https://doi.org/10.1016/j.canlet.2008.02.027>
- Paulech et al., 2013** – Paulech, J., Solis, N., Edwards, A.V., Puckeridge, M., White, M.Y., Cordwell, S.J. (2013). Large-scale capture of peptides containing reversibly oxidized cysteines by thiol-disulfide exchange applied to the myocardial redox proteome. *Analytical chemistry*. 85(7): 3774-3780. DOI: <https://doi.org/10.1021/ac400166e>
- Pena et al., 2022** – Pena, I., Pena-Vina, E., Rodriguez-Avial, I., Picazo, J.J., Gómez-González, Á., Culebras, E. (2022). Comparison of performance of MALDI-TOF MS and MLST for biotyping carbapenemase-producing Klebsiella pneumoniae sequence types ST11 and ST101 isolates. *Enfermedades infecciosas y microbiología clínica (English ed.)*. 40(4): 172-178. DOI: <https://doi.org/10.1016/j.eimce.2020.10.011>
- Person et al., 2003** – Person, M.D., Monks, T.J., Lau, S.S. (2003). An integrated approach to identifying chemically induced posttranslational modifications using comparative MALDI-MS and targeted HPLC-ESI-MS/MS. *Chemical research in toxicology*. 16(5): 598-608. DOI: <https://doi.org/10.1021/tx020109f>

- Phulara, Seneviratne, 2024** – Phulara, N.R., Seneviratne, H.K. (2024). Mass spectrometry imaging-based multi-omics approaches to understand drug metabolism and disposition. *Journal of Mass Spectrometry*. 59(7): e5042. DOI: <https://doi.org/10.1002/jms.5042>
- Pliss et al., 2021** – Pliss, A., Kuzmin, A.N., Lita, A., Kumar, R., Celiku, O., Atilla-Gokcumen, G.E., Gokcumen, O., Chandra, D., Larion, M., Prasad, P.N. (2021). A single-organelle optical omics platform for cell science and biomarker discovery. *Analytical chemistry*. 93(23): 8281-8290. DOI: <https://doi.org/10.1021/acs.analchem.1c01131>
- Polikar, Bessi, 1970** – Polikar, A., Bessi, M. (1970). Elementy patologii kletki [Elements of cell pathology]. M.: Mir. [in Russian]
- Portugalov et al., 1964** – Portugalov, V.V., Krasnov, I.B., Ball', T.V. (1964). A histochemical determination of the denaturation of the proteins of the nerve cell induced by alcohol. *Bulletin of Experimental Biology and Medicine*. 55(4): 458-460. DOI: <https://doi.org/10.1007/BF00785679>
- Pralea et al., 2020** – Pralea, I.E., Moldovan, R.C., Tigu, A.B., Ionescu, C., Iuga, C.A. (2020). Mass spectrometry-based omics for the characterization of triple-negative breast cancer bio-signature. *Journal of personalized medicine*. 10(4): 277. DOI: <https://doi.org/10.3390/jpm10040277>
- Pranada et al., 2016** – Pranada, A.B., Schwarz, G., Kostrzewa, M. (2016). MALDI Biotyping for microorganism identification in clinical microbiology. *Advances in MALDI and laser-induced soft ionization mass spectrometry*, 197-225. DOI: [10.1007/978-3-319-04819-2_11](https://doi.org/10.1007/978-3-319-04819-2_11)
- Quanico et al., 2017** – Quanico, J., Franck, J., Wisztorski, M., Salzet, M., Fournier, I. (2017). Integrated mass spectrometry imaging and omics workflows on the same tissue section using grid-aided, parafilm-assisted microdissection. *Biochimica et Biophysica Acta (BBA)-General Subjects*. 1861(7): 1702-1714. DOI: <https://doi.org/10.1016/j.bbagen.2017.03.006>
- Quintá et al., 2016** – Quintá, H.R., Wilson, C., Blidner, A.G., González-Billault, C., Pasquini, L.A., Rabinovich, G.A., Pasquini, J.M. (2016). Ligand-mediated Galectin-1 endocytosis prevents intraneuronal H₂O₂ production promoting F-actin dynamics reactivation and axonal re-growth. *Experimental neurology*. 283: 165-178. DOI: <https://doi.org/10.1016/j.expneurol.2016.06.009>
- Ralph et al., 2010** – Ralph, S.J., Rodríguez-Enríquez, S., Neuzil, J., Saavedra, E., Moreno-Sánchez, R. (2010). The causes of cancer revisited: “mitochondrial malignancy” and ROS-induced oncogenic transformation—why mitochondria are targets for cancer therapy. *Molecular aspects of medicine*. 31(2): 145-170. DOI: <https://doi.org/10.1016/j.mam.2010.02.008>
- Rezende et al., 2018** – Rezende, F., Brandes, R.P., Schröder, K. (2018). Detection of hydrogen peroxide with fluorescent dyes. *Antioxidants & Redox Signaling*. 29(6): 585-602. DOI: <https://doi.org/10.1089/ars.2017.7401>
- Rhee et al., 2010** – Rhee, S.G., Chang, T.S., Jeong, W., Kang, D. (2010). Methods for detection and measurement of hydrogen peroxide inside and outside of cells. *Molecules and cells*. 29: 539-549. DOI: <https://doi.org/10.1007/s10059-010-0082-3>
- Ryabchykov et al., 2018** – Ryabchykov, O., Popp, J., Bocklitz, T. (2018). Fusion of MALDI spectrometric imaging and Raman spectroscopic data for the analysis of biological samples. *Frontiers in Chemistry*. 6: 257. DOI: <https://doi.org/10.3389/fchem.2018.00257>
- Sadri et al., 2022** – Sadri, H., Aghaei, M., Akbari, V. (2022). Nisin induces apoptosis in cervical cancer cells via reactive oxygen species generation and mitochondrial membrane potential changes. *Biochemistry and Cell Biology*. 100(2): 136-141. DOI: <https://doi.org/10.1139/bcb-2021-0225>
- Samoylenko et al., 2013** – Samoylenko, A., Hossain, J.A., Mennerich, D., Kellokumpu, S., Hiltunen, J.K., Kietzmann, T. (2013). Nutritional countermeasures targeting reactive oxygen species in cancer: from mechanisms to biomarkers and clinical evidence. *Antioxidants & redox signaling*. 19(17): 2157-2196. DOI: <https://doi.org/10.1089/ars.2012.4662>
- Sanders, Edwards, 2020** – Sanders K.L., Edwards J.L. (2020). Nano-liquid chromatography-mass spectrometry and recent applications in omics investigations. *Analytical Methods*. 12(36): 4404-4417. DOI: <https://doi.org/10.1039/doay01194k>
- Schmitt et al., 2023** – Schmitt, R., Qayum, S., Pliss, A., Kuzmin, A.N., Muthaiah, V.P.K., Kaliyappan, K., Prasad, P.N., Mahajan, S.D. (2023). Mitochondrial Dysfunction and Apoptosis in Brain Microvascular Endothelial Cells Following Blast Traumatic Brain Injury. *Cellular and Molecular Neurobiology*. 43(7): 3639-3651. DOI: <https://doi.org/10.1007/s10571-023-01372-2>
- Schulte-Hermann et al., 2006** – Schulte-Hermann, R., Teufelhofer, O., Parzefall, W., Freiler, C., Grasl-Kraupp, B., Gerner, C. (2006). Reactive oxygen from macrophages and chemical carcinogenesis. *Toxicology Letters*. (164): S4. DOI: <http://dx.doi.org/10.1016/j.toxlet.2006.06.012>

Shen et al., 2021 – *Shen, Y., Yue, J., Xu, W., Xu, S.* (2021). Recent progress of surface-enhanced Raman spectroscopy for subcellular compartment analysis. *Theranostics*. 11(10): 4872. DOI: <https://doi.org/10.7150/thno.56409>

Shi et al., 1998 – *Shi, X., Castranova, V., Halliwell, B., Vallyathan, V.* (1998). Reactive oxygen species and silica-induced carcinogenesis. *Journal of Toxicology and Environmental Health, Part B Critical Reviews*. 1(3): 181-197. DOI: <https://doi.org/10.1080/10937409809524551>

Shimura, Ushiyama, 2024 – *Shimura, T., Ushiyama, A.* (2024). Mitochondrial reactive oxygen species-mediated fibroblast activation has a role in tumor microenvironment formation in radiation carcinogenesis. *Radiation Protection Dosimetry*. 200(16-18): 1590-1593. DOI: <https://doi.org/10.1093/rpd/ncae027>

Shkarina et al., 1984 – *Shkarina, T.N., Zatsepina, G.N., Kasatkina, V.V., Kozinets, G.I., Tarasova, I.M.* (1984). Electrical charge of the surface of lymphocytes and their capacity for mitotic division in induced carcinogenesis. *Biophysics*. 29(1): 102-106.

Siddhanta et al., 2023 – *Siddhanta, S., Kuzmin, A.N., Pliss, A., Baev, A.S., Khare, S.K., Chowdhury, P.K., Ganguli, A.K., Prasad, P.N.* (2023). Advances in Raman spectroscopy and imaging for biomedical research. *Advances in Optics and Photonics*. 15(2): 318-384. DOI: <http://dx.doi.org/10.1364/AOP.479884>

Skates, 2022 – *Skates, E.* (2022). Measurement and manipulation of mitochondrial membrane potential in cancer cells (Doctoral dissertation, University of Warwick).

Skottvoll, 2022 – *Skottvoll, F.S.* (2022). *Liver organoids, mass spectrometry, and separation science (Dissertation for the degree Philosophiae Doctor)*. University of Oslo (Department of Chemistry; Faculty of Mathematics and Natural Sciences). DOI: <https://doi.org/10.1002/anse.202100051>

Smets et al., 2021 – *Smets, T., De Keyser, T., Tousseyen, T., Waelkens, E., De Moor, B.* (2021). Correspondence-aware manifold learning for microscopic and spatial omics imaging: a novel data fusion method bringing mass spectrometry imaging to a cellular resolution. *Analytical Chemistry*. 93(7): 3452-3460. DOI: <https://doi.org/10.1021/acs.analchem.oco4759>

Smith et al., 2014 – *Smith, M.R., Zhou, F., Kumar, P.V., Beggs, R., Velu, S., Landar, A., Murphy, M.* (2014). Metabolic Reprogramming by a Mitochondria-Targeted Electrophile in Breast Cancer Cells. *Free Radical Biology and Medicine*. 76: S132. DOI: <http://dx.doi.org/10.1016/j.freeradbiomed.2014.10.212>

Smolyarova et al., 2022 – *Smolyarova, D.D., Podgorny, O.V., Bilan, D.S., Belousov, V.V.* (2022). A guide to genetically encoded tools for the study of H₂O₂. *The FEBS journal*. 289(18): 5382-5395. DOI: <https://doi.org/10.1111/febs.16088>

Somboro et al., 2014 – *Somboro, A.M., Essack, S.Y., Tiwari, D., Shobo, A., Bester, L.A., Kruger, H.G., Govender, T.* (2014). Evaluation of MALDI Biotyping for Rapid Subspecies Identification of Carbapenemase-Producing Bacteria via Protein Profiling. *Mass Spectrometry Letters*. 5(4): 110-114. DOI: <http://dx.doi.org/10.5478/MSL.2014.5.4.110>

Standeven, Wetterhahn, 1991 – *Standeven, A.M., Wetterhahn, K.E.* (1991). Is there a role for reactive oxygen species in the mechanism of chromium (VI) carcinogenesis? *Chemical research in toxicology*. 4(6): 616-625. DOI: <https://doi.org/10.1021/tx00024a003>

Tamura et al., 2013 – *Tamura, M., Mutoh, M., Fujii, G., Matsui, H.* (2013). Involvement of Mitochondrial Reactive Oxygen Species in Gastric Carcinogenesis. *J Gastroint Dig Syst*. 3: 150. DOI: <https://doi.org/10.4172/2161-069X.1000150>

Tian et al., 2021 – *Tian, Y., Xu, W., Ma, K., Cong, L., Shen, Y., Han, X., Liang, C., Liang, L., Qi, G., Jin, Y., Xu, S.* (2021). Label-free analysis of cell membrane proteins via evanescent field excited surface-enhanced Raman scattering. *The Journal of Physical Chemistry Letters*. 12(43): 10720-10727. DOI: <https://doi.org/10.1021/acs.jpclett.1c02966>

Tokuoka, Morioka, 1957 – *Tokuoka, S., Morioka, H.* (1957). The membrane potential of the human cancer and related cells (I). *Gann*. 48(4): 353-354.

Valavanidis, 1994 – *Valavanidis, A.* (1994). Ultraviolet radiation and skin cancer implication of free radical reactions and reactive oxygen species in skin carcinogenesis. *Review of Clinical Pharmacology and Pharmacokinetics (International Edition)*. 8: 101. DOI: <https://doi.org/10.1042/bss0610047>

Valavanidis, 2019 – *Valavanidis, A.* (2019). Oxidative stress and pulmonary carcinogenesis through mechanisms of reactive oxygen species. How respirable particulate matter, fibrous dusts,

and ozone cause pulmonary inflammation and initiate lung carcinogenesis. *Oxidative Stress in Lung Diseases*. 1: 247-265. DOI: https://doi.org/10.1007/978-981-13-8413-4_13

Vostrikova et al., 2020 – Vostrikova, S.M., Grinev, A.B., Gogvadze, V.G. (2020). Reactive oxygen species and antioxidants in carcinogenesis and tumor therapy. *Biochemistry (Moscow)*. 85: 1254-1266. DOI: <https://doi.org/10.1134/s0006297920100132>

Wang et al., 2023a – Wang, X., Han, J., Li, Z., Li, B., Wan, Y., Liu, L. (2023). Insight into plant spatial omics: mass spectrometry imaging. *Frontiers in Plant Science*. 14: 1273010. DOI: <https://doi.org/10.3389/fpls.2023.1273010>

Wang et al., 2022 – Wang, J., Pursell, M.E., DeVor, A., Awoyemi, O., Valentine, S.J., Li, P. (2022). Portable mass spectrometry system: instrumentation, applications, and path to ‘omics analysis. *Proteomics*. 22(23-24): 2200112. DOI: <https://doi.org/10.1002/pmic.202200112>

Wang et al., 2023b – Wang, Z., Liu, B., Lin, L., Qiao, L. (2023). Mass spectrometry for mitochondrial multi-omics. *TrAC Trends in Analytical Chemistry*. 163: 117063. DOI: <https://doi.org/10.1016/j.trac.2023.117063>

Wang, 2009 – Wang, G. (2009). NADPH oxidase and reactive oxygen species as signaling molecules in carcinogenesis. *Frontiers of Medicine in China*. 3: 1-7. DOI: <https://doi.org/10.1007/s11684-009-0018-5>

Weller et al., 2014 – Weller, J., Kizina, K.M., Can, K., Bao, G., Müller, M. (2014). Response properties of the genetically encoded optical H₂O₂ sensor HyPer. *Free Radical Biology and Medicine*. 76: 227-241. DOI: <https://doi.org/10.1016/j.freeradbiomed.2014.07.045>

Wolyniak et al., 2018 – Wolyniak, M.J., Reyna, N.S., Plymale, R., Pope, W.H., Westholm, D.E. (2018). Mass spectrometry as a tool to enhance “-omics” education. *Journal of Microbiology & Biology Education*. 19(1): 10-1128. DOI: <https://doi.org/10.1128/jmbe.v19i1.1459>

Wu, Ni, 2015 – Wu, Q., Ni, X. (2015). ROS-mediated DNA methylation pattern alterations in carcinogenesis. *Current drug targets*. 16(1): 13-19. DOI: <https://doi.org/10.2174/1389450116666150113121054>

Yang, Brackenbury, 2013 – Yang, M., Brackenbury, W.J. (2013). Membrane potential and cancer progression. *Frontiers in physiology*. 4: 185. DOI: <https://doi.org/10.3389/fphys.2013.00185>

Yang et al., 2023 – Yang, E., Kim, J.H., Tressler, C.M., Shen, X.E., Brown, D.R., Johnson, C.C., Hahm, T.H., Barman, I., Glunde, K. (2023). RaMALDI: enabling simultaneous Raman and MALDI imaging of the same tissue section. *Biosensors and Bioelectronics*. 239: 115597. DOI: <https://doi.org/10.1016/j.bios.2023.115597>

Yang, 2014 – Yang, Y. (2014). Alternative Approaches to Optical Sensing of the Redox State. In *Natural Biomarkers for Cellular Metabolism* (pp. 208-229). CRC Press. DOI: <http://dx.doi.org/10.1201/b17427-11>

Ye et al., 2011 – Ye, X.Q., Wang, G.H., Huang, G.J., Bian, X.W., Qian, G.S., Yu, S.C. (2011). Heterogeneity of mitochondrial membrane potential: a novel tool to isolate and identify cancer stem cells from a tumor mass?. *Stem Cell Reviews and Reports*. 7: 153-160. DOI: <https://doi.org/10.1007/s12015-010-9122-9>

Zaikin, Borisov, 2021 – Zaikin, V.G., Borisov, R.S. (2021). Mass spectrometry as a crucial analytical basis for omics sciences. *Journal of Analytical Chemistry*. 76: 1567-1587. DOI: <https://doi.org/10.1134/S1061934821140094>

Zhang, Qiao, 2024 – Zhang, D., Qiao, L. (2024). Microfluidics coupled mass spectrometry for single cell multi-omics. *Small Methods*. 8(1): 2301179. DOI: <https://doi.org/10.1002/smtd.202301179>

Zhang et al., 2007 – Zhang, X., Wie, D., Yap, Y., Li, L., Guo, S., Chen, F. (2007). Mass spectrometry-based “omics” technologies in cancer diagnostics. *Mass spectrometry reviews*. 26(3): 403-431. DOI: <https://doi.org/10.1002/mas.20132>

Zhang et al., 2019 – Zhang, P., Wang, L., Fang, Y., Zheng, D., Lin, T., Wang, H. (2019). Label-free exosomal detection and classification in rapid discriminating different cancer types based on specific Raman phenotypes and multivariate statistical analysis. *Molecules*. 24(16): 2947. DOI: <https://doi.org/10.3390/molecules24162947>

Zhang et al., 2023 – Zhang, H., Delafield, D.G., Li, L. (2023). Mass spectrometry imaging: the rise of spatially resolved single-cell omics. *Nature Methods*. 20(3): 327-330. DOI: <https://doi.org/10.1038/s41592-023-01774-6>

Zhao, Cai, 2023 – Zhao, C., Cai, Z. (2023). Mass spectrometry-based omics and imaging technique: a novel tool for molecular toxicology and health impacts. *Reviews of Environmental Contamination and Toxicology*. 261(1): 10. DOI: <http://dx.doi.org/10.1007/s44169-023-00032-2>

Zhao et al., 2022 – Zhao, C., Dong, J., Deng, L., Tan, Y., Jiang, W., Cai, Z. (2022). Molecular network strategy in multi-omics and mass spectrometry imaging. *Current Opinion in Chemical Biology*. 70: 102199. DOI: <https://doi.org/10.1016/j.cbpa.2022.102199>

Zhao et al., 2023 – Zhao, P., Feng, Y., Wu, J., Zhu, J., Yang, J., Ma, X., Ouyang, Z., Zhang, X., Zhang, W., Wang, W. (2023). Efficient sample preparation system for multi-omics analysis via single cell mass spectrometry. *Analytical Chemistry*. 95(18): 7212-7219. DOI: <https://doi.org/10.1021/acs.analchem.2c05728>

Ziech et al., 2011 – Ziech, D., Franco, R., Pappa, A., Panayiotidis, M.I. (2011). Reactive Oxygen Species (ROS) – Induced genetic and epigenetic alterations in human carcinogenesis. *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis*. 711(1-2): 167-173. DOI: <https://doi.org/10.1016/j.mrfmmm.2011.02.0150>

Ziech et al., 2012 – Ziech, D., Anestopoulos, I., Hanafi, R., Voulgaridou, G.P., Franco, R., Georgakilas, A.G., Pappa, A., Panayiotidis, M.I. (2012). Pleiotrophic effects of natural products in ROS-induced carcinogenesis: the role of plant-derived natural products in oral cancer chemoprevention. *Cancer Letters*. 327(1-2): 16-25. DOI: <https://doi.org/10.1016/j.canlet.2012.02.025>

Ретроспективный анализ схем раманомики: от количественной раманомики с использованием глубинных сверточных нейронных сетей для point-of-care-диагностики до молекулярно-оптических лазерных анализаторов. Часть 1 (Библиографический обзор)

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Аннотация. В данной обзорной работе производится ретроспективный анализ технологий раманомики и её методологических предшественников, направленный от современной количественной раманомики с использованием глубоких сверточных нейронных сетей (используемой для интраоперационной диагностики и point-of-care-диагностики) до Molecular Optical Laser Examiners (MOLE) 1970-х гг. Первая часть обзора, публикуемая в настоящем выпуске, рассматривает современные направления данного тренда, в то время как во второй части представляются достижения более раннего периода. В первой части внимание уделяется приложениям раманомики для диагностики супрамолекулярных патологий, механизмов апоптоза, парабиоза, онкогенеза, ряда редокс-патологий (а также эффектов воздействия активных форм кислорода на клетки и ткани), повреждений гематоэнцефалического барьера и нейротравм, затрагивающих цитоархитектонику мозга и, шире, архитектуру нейрональных коннектомов. Указывается ряд работ, позволяющих говорить о рамановском анализе в задачах спектральной сравнительно-патологической органеллографии цитоплазмы. Приводятся сведения об интегрируемости раманомики и методов масс-спектрометрического картирования или RaMALDI, в том числе – для задач MALDI-биотайпинга (как правило, используемого в клинической микробиологии).

Ключевые слова: раманомика, количественная раманомика, спектраломика, омиксный ультраструктурный анализ, MALDI-имэджинг, RaMALDI, синхронизированные MALDI-имэджинг и рамановская визуализация, безметочный времязадержанный мониторинг на уровне одиночных клеток, интраоперационная диагностика, РОС-диагностика, сверточные нейросети.

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